

**How Difficult is it to Eliminate Coronavirus From Surfaces?**  
**Prof. Jean-Yves Maillard, Cardiff University, Wales**  
**A Webber Training Teleclass**



**CARDIFF UNIVERSITY**  
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**HOW DIFFICULT IS IT TO ELIMINATE CORONAVIRUS FROM SURFACES?**

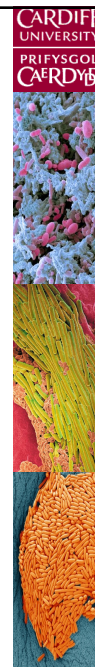
**Jean-Yves Maillard**  
Cardiff School of Pharmacy and Pharmaceutical Sciences  
Cardiff University

Hosted by Dr. Lynne Schulster

[www.webbertraining.com](http://www.webbertraining.com) June 11, 2020

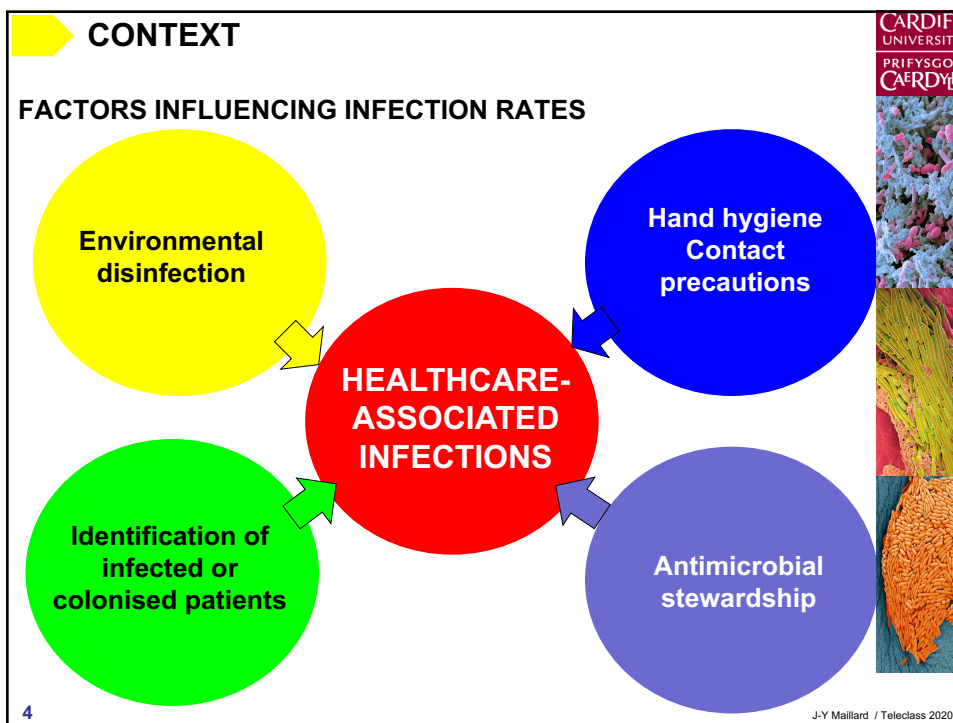
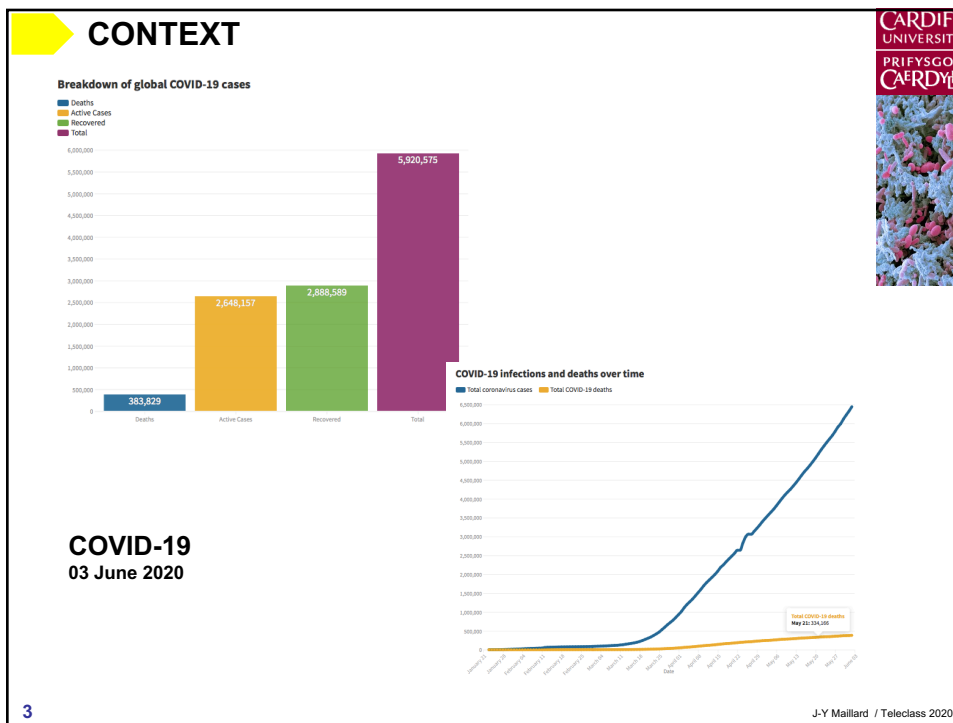
**OVERVIEW**

-  **Context**
-  **Role of surfaces in pathogen transmission**
-  **Virus structure & impact on susceptibility**
-  **Virus susceptibility to chemical disinfection**
-  **Conclusions**



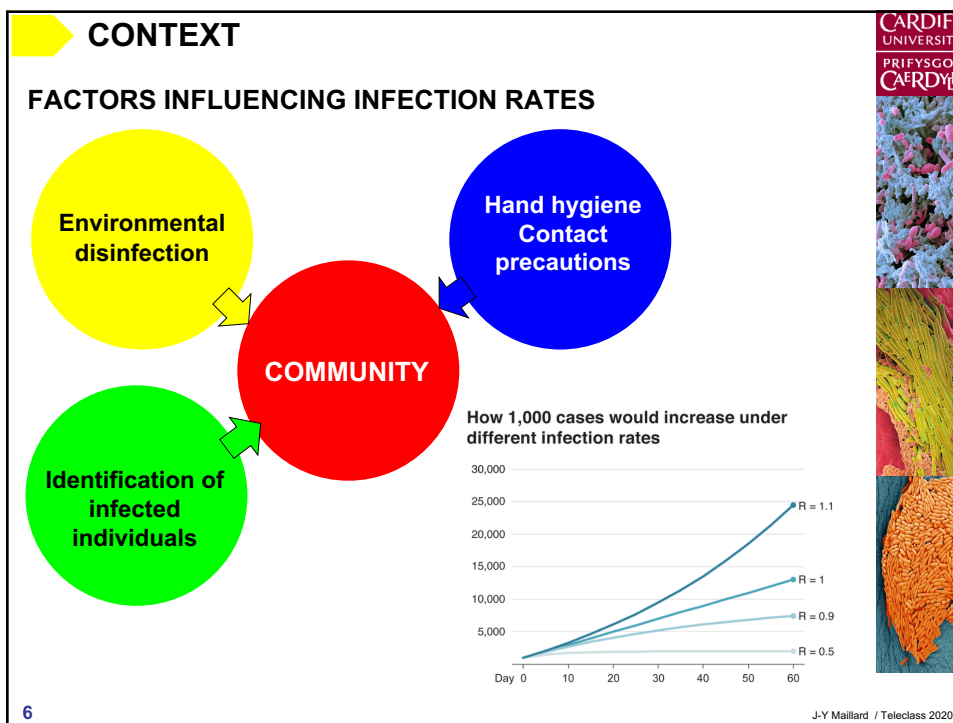
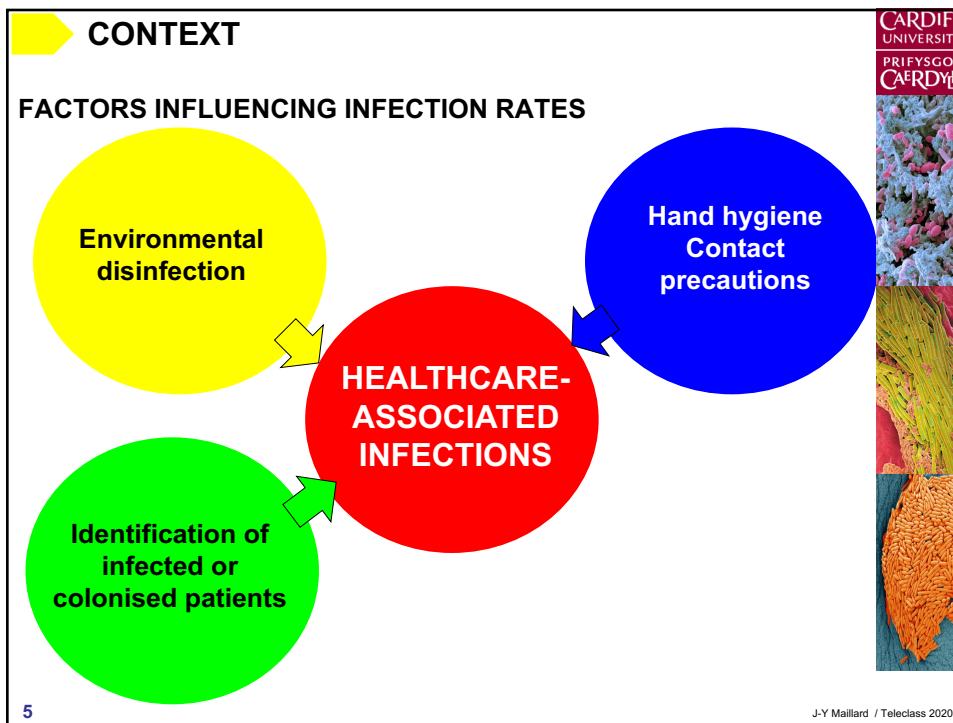
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**CONTEXT**

**Pathogen transmission**

• Large droplets ( $>100\ \mu\text{m}$ ): Fast deposition due to the domination of gravitational force  
 • Medium droplets between 5 and  $100\ \mu\text{m}$   
 • Small droplets or droplet nuclei, or aerosols ( $< 5\ \mu\text{m}$ ): Responsible for airborne transmission

Fig 4. Illustration of different transmission routes. Small droplets ( $<5\ \mu\text{m}$ ), sometimes called aerosols, are responsible for the short-range airborne route, long-range airborne route, and indirect contact route; large droplets are responsible for the direct spray route and indirect contact route.

Wei & Li. *Am J Infect Control*, 2016; 44: S102-S108.

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**CONTEXT**

**Pathogen transmission in hospital**

Isolation room: Puff expiratory airflow, Wake flow  
 Anteroom: Vortices causing mixing, Two-way airflow  
 Corridor: Door vortices, Body thermal plume, Warm air, Cold air

Wei & Li. *Am J Infect Control*, 2016; 44: S102-S108.

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**CONTEXT**

**Pathogen transmission in hospital**

- Coughing: 24-23,600 droplets / 1L exhaled air
- Talking: 4-600 droplets / 1L exhaled air
- Breathing by mouth: 1-320 droplets / 1L exhaled air

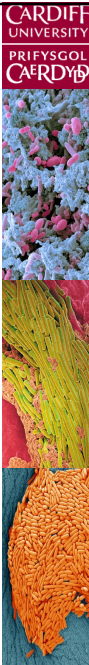
➤ People in close contact can contaminate each other by direct spray from large droplets or by inhalation of smaller droplets.

➤ Contamination of the other person's skin or clothing can lead to self-inoculation through indirect contact transmission.


Wei & Li. Am J Infect Control, 2016; 44: S102-S108.

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**ROLE OF SURFACES IN MICROBIAL TRANSMISSION**



High touch surfaces

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## ▶ ROLE OF SURFACES IN MICROBIAL TRANSMISSION

**Microorganisms survival on surfaces proximal to patients (high-touch surfaces)**

**Low infectious dose for some pathogens**

**Pathogens survival on surfaces at concentration sufficient for transmission**

**Genotypic link between bacteria isolated from patients and surfaces**

**1970s - 1990s: THE DARK AGES: AN ALMOST COMPLETE DENIAL OF SURFACE CONTAMINANTS!**

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## ▶ ROLE OF SURFACES IN MICROBIAL TRANSMISSION

### Surfaces touched by staff

Med-Surg unit (touches per visit)

- Bedrails = 3.1
- Over-bed table = 1.6
- IV pump = 1.4
- Bed surface = 1.3

- Average surfaces per interaction
  - ICU = 44, Med-Surg = 15
- Combining the data:
  - 82 people per day visit a typical patient room
  - 2/3 are clinical staff. 1/3 are not.
  - Bedrails get touched up to 256 times per day

Huslage et al. Infect Cont and Hosp Epidemiol, 2010;31: 850-3.

Surface	Mean Number of Surface Contacts per Transmission
Bed rails	3.1
Over-bed table	1.6
IV pump	1.4
Bed surface	1.3
Other surfaces	< 1.0

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## ROLE OF SURFACES IN MICROBIAL TRANSMISSION

### Surfaces touched by visitors

Visitors contact a range of surfaces/objects

- 33.5% = contact with the environment only (Most common level of touch)
- 27.1% = patient's intact skin
- 17.8% = blood or body fluids
- 16.0% = the person touched nothing in the room

- Nonclinical visitors and personal visitors were exposed to blood or body fluids 19% of the time
- 75% of the time, nonclinical visitors touch environmental surfaces only.

Care Role	Blood or body fluids	Intact skin only	Environment only	Nothing in room
Nursing (N=1,413)	15.6	34.6	20.2	29.7
Medical (N=642)	37.6	20.5	10.3	31
Other clinical (N=140)	21.4	20.7	31.4	26.4
Nonclinical (N=245)	19.2	74.7	6.1	0
Visitors (N=723)	19.6	28.5	45.1	8.8

**Figure 1.** Contact with patient environment only was the most common level of touch (33.5% of visits), followed by patient's intact skin (27.1%) and patient's blood or body fluids (17.8%).

Cohen et al. J Comm J Qual Patient Safety, 2012;38:560-5. J-Y Maillard / Teleclass 2020

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## ROLE OF SURFACES IN MICROBIAL TRANSMISSION

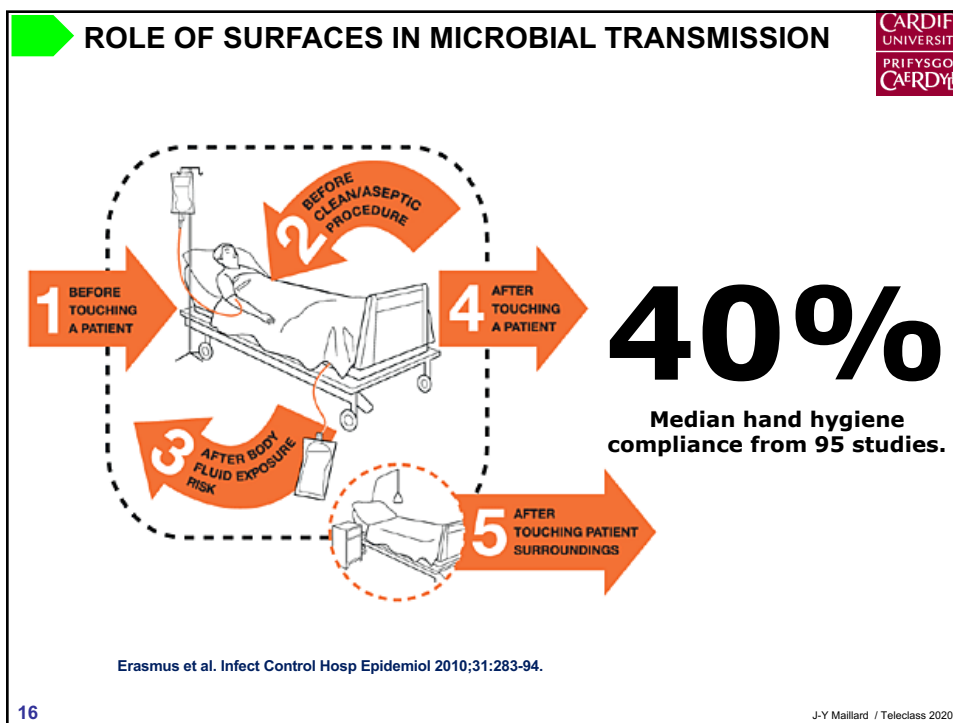
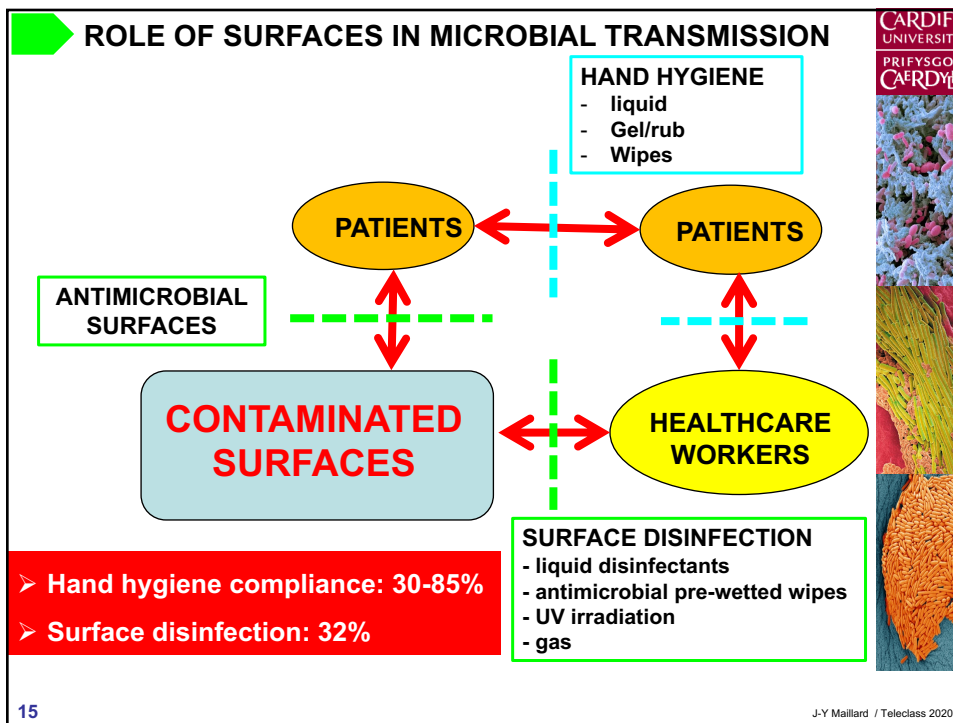
Observations	Hospital 1	Hospital 2
% observations where staff washed hands	28	20
% observations where staff used alcoholic hand rub	30	9
Of those incidences where no gloves worn, % incidences where staff used alcoholic hand rub	41	14
% staff wearing no gloves and used no AHR, but washed hands	17	19
% staff using no protection/skin sanitisation	19	46
<b>% potential staff to object cross- contamination</b>	<b>30</b>	<b>59</b>
% potential staff to patient cross-contamination	4	0
<b>% potential object to object cross- contamination</b>	<b>70</b>	<b>88</b>
% potential object to patient cross-contamination	20	9
<b>% potential patient to object cross-contamination</b>	<b>17</b>	<b>9</b>

Low frequency of hand sanitisation, particularly with use of AHR lead to high incidence of potential cross contamination

Cheeseman et al. J Hosp Infect, 2009; 72: 319-25. J-Y Maillard / Teleclass 2020

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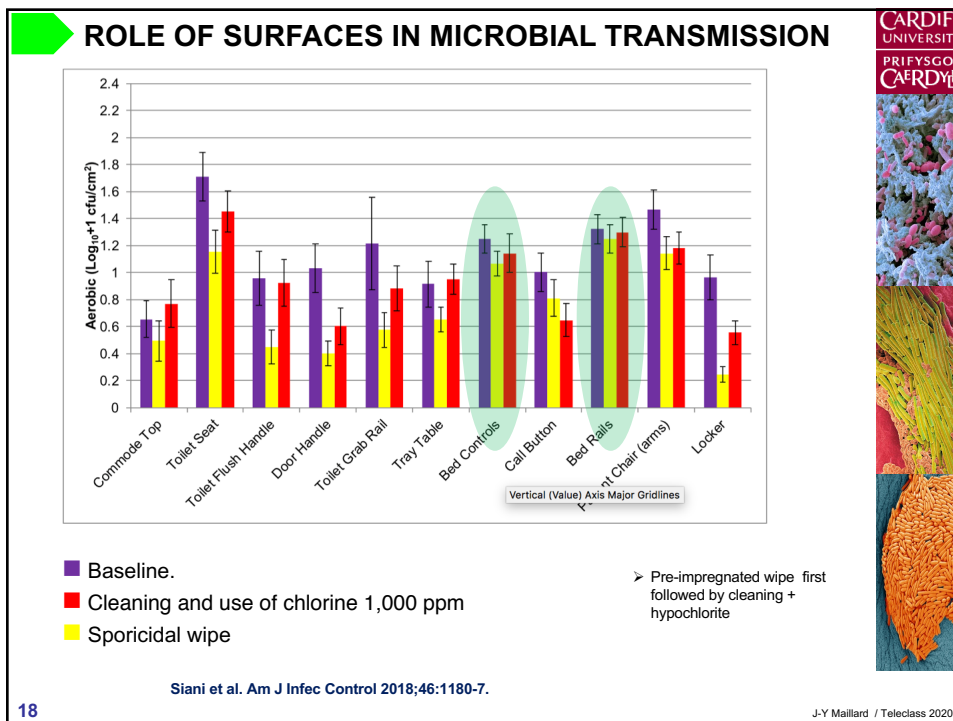
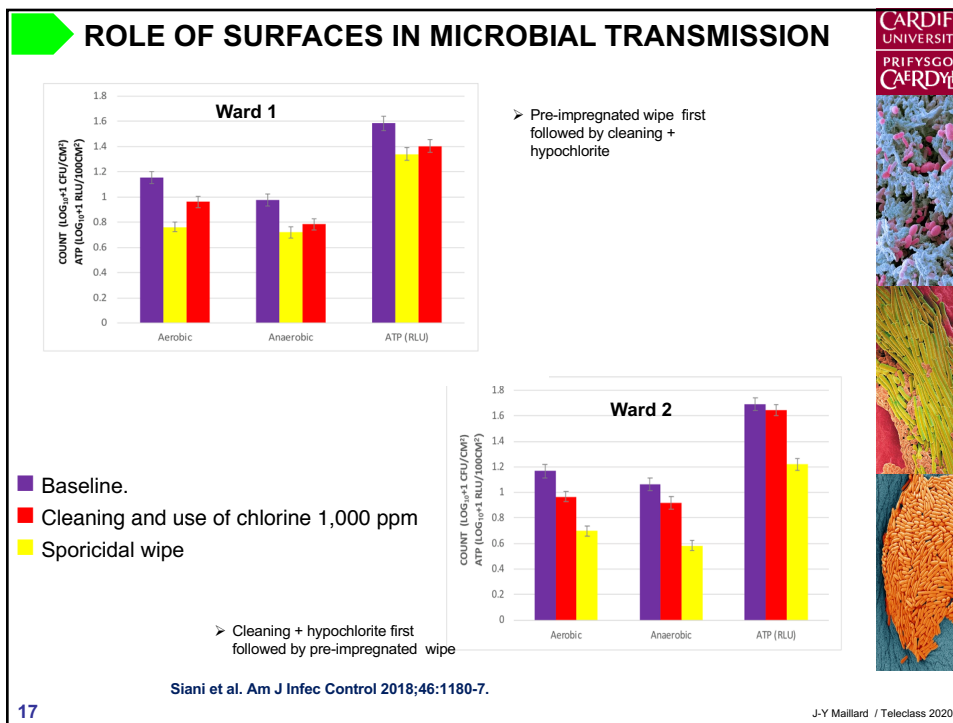
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▶

ROLE OF SURFACES IN MICROBIAL TRANSMISSION




### Survival of pathogens on hospital surfaces

PATHOGEN	SURIVAL TIME
<i>S. aureus</i> (incl. MRSA)	7 days to >12 months
<i>Enterococcus</i> spp. (incl VRE)	5 days to >46 months
<i>Acinetobacter</i> spp.	3 days to 11 months
<i>C. difficile</i> (spores)	> 5 months
<b>Norovirus (&amp; feline calicivirus)</b>	<b>8 h to &gt; 2 weeks</b>
<i>Ps. aeruginosa</i>	6 h to 16 months
<i>Klebsiella</i> spp.	2h to 30 months


Hota et al. *Clin Infect Dis* 2004;39:1182-9  
Kramer et al. *BMC Infect Dis* 2006;6:130

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▶

ROLE OF SURFACES IN MICROBIAL TRANSMISSION




### Persistence of human coronavirus and coronavirus surrogates Reviewed by Kampf et al. *J Hosp Infect* 2020;104:246-51.

Persistence of coronaviruses on different types of inanimate surfaces

Type of surface	Virus	Strain / isolate	Inoculum (viral titer)	Temperature	Persistence
Steel	MERS-CoV	Isolate HCoV-EMC/2012	10 <sup>5</sup>	20°C	48 h
				30°C	8–24 h
	TGEV	Unknown	10 <sup>6</sup>	4°C	≥ 28 d
				20°C	3–28 d
	MHV	Unknown	10 <sup>6</sup>	40°C	4–96 h
				4°C	≥ 28 d
Aluminium	HCoV	Strain 229E	10 <sup>3</sup>	20°C	4–28 d
				40°C	4–96 h
Metal	HCoV	Strains 229E and OC43	5 x 10 <sup>3</sup>	21°C	5 d
				RT	2–8 h
Wood	SARS-CoV	Strain P9	10 <sup>5</sup>	RT	5 d
				RT	4 d
Paper	SARS-CoV	Strain P9	10 <sup>5</sup>	RT	4–5 d
				RT	24 h
Glass	SARS-CoV	Strain GUV6109	10 <sup>6</sup>	RT	3 h
				RT	< 5 min
	SARS-CoV	Strain P9	10 <sup>5</sup>	RT	4 d
					4 d
	HCoV	Strain 229E	10 <sup>3</sup>	21°C	5 d
					5 d
Plastic	SARS-CoV	Strain HKU39849	10 <sup>5</sup>	22°-25°C	≤ 5 d
				20°C	48 h
PVC	MERS-CoV	Isolate HCoV-EMC/2012	10 <sup>5</sup>	30°C	8–24 h
				30°C	8–24 h
	SARS-CoV	Strain P9	10 <sup>5</sup>	RT	4 d
					4 d
	SARS-CoV	Strain FFM1	10 <sup>7</sup>	RT	6–9 d
					6–9 d
HCoV	Strain 229E	10 <sup>7</sup>	RT	2–6 d	
				2–6 d	
Silicon rubber	HCoV	Strain 229E	10 <sup>3</sup>	21°C	5 d
				21°C	5 d
Surgical glove (latex)	HCoV	Strains 229E and OC43	5 x 10 <sup>3</sup>	21°C	≤ 8 h
				21°C	≤ 8 h
Disposable gown	SARS-CoV	Strain GUV6109	10 <sup>6</sup>	RT	2 d
				RT	24 h
Ceramic	HCoV	Strain 229E	10 <sup>4</sup>	RT	1 h
				RT	1 h
Teflon	HCoV	Strain 229E	10 <sup>3</sup>	21°C	5 d
				21°C	5 d

MERS = Middle East Respiratory Syndrome; HCoV = human coronavirus; TGEV = transmissible gastroenteritis virus; MHV = mouse hej  
SARS = Severe Acute Respiratory Syndrome; RT = room temperature.

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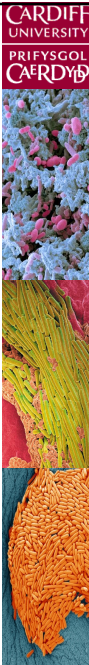
## ▶ ROLE OF SURFACES IN MICROBIAL TRANSMISSION

### Human coronavirus- SARS-CoV Lai et al. Clin Infect Dis 2005;41:e67

- ▶ Simulates a situation in which large respiratory droplets (volume, 5 mL; radius, ~1 mm) that contain the virus fall onto paper

Inoculation, TCID <sub>50</sub> /mL	Time taken to inactivate SARS-CoV, by surface		
	Paper	Disposable gown	Cotton gown
10 <sup>6</sup>	24 h	2 days	24 h
10 <sup>5</sup>	3 h	24 h	1 h
10 <sup>4</sup>	<5 min	1 h	5 min

- ▶ “Even with a higher concentration of virus (10<sup>4</sup> TCID<sub>50</sub>/mL) than would normally occur in NPA samples (10<sup>2.2</sup> TCID<sub>50</sub>/mL), no virus infectivity remained after the paper was dried.”
- ▶ “Paper contaminated with a higher concentration of virus (equivalent to that of fecal excreta (i.e., 10<sup>5</sup> TCID<sub>50</sub>/mL) was not infectious after 3 h, and no viral infectivity was shown after 24 h, even with a concentration of 10<sup>6</sup> TCID<sub>50</sub>/mL.”



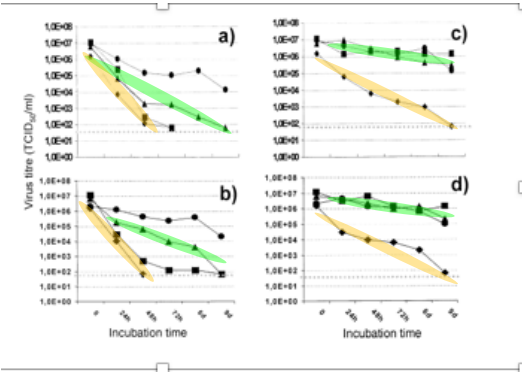
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## ▶ ROLE OF SURFACES IN MICROBIAL TRANSMISSION

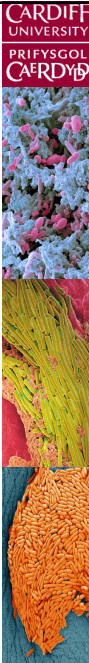
### Stability of SARS-CoV Rabenau et al. Med Microbiol Immunol 2005;194:1-6.

a,b: surface  
c,d: suspension  
b,d: organic load

● adenovirus type-3  
▲ SARS-CoV  
◆ h-CoV (229E)  
■ □ HSV-1



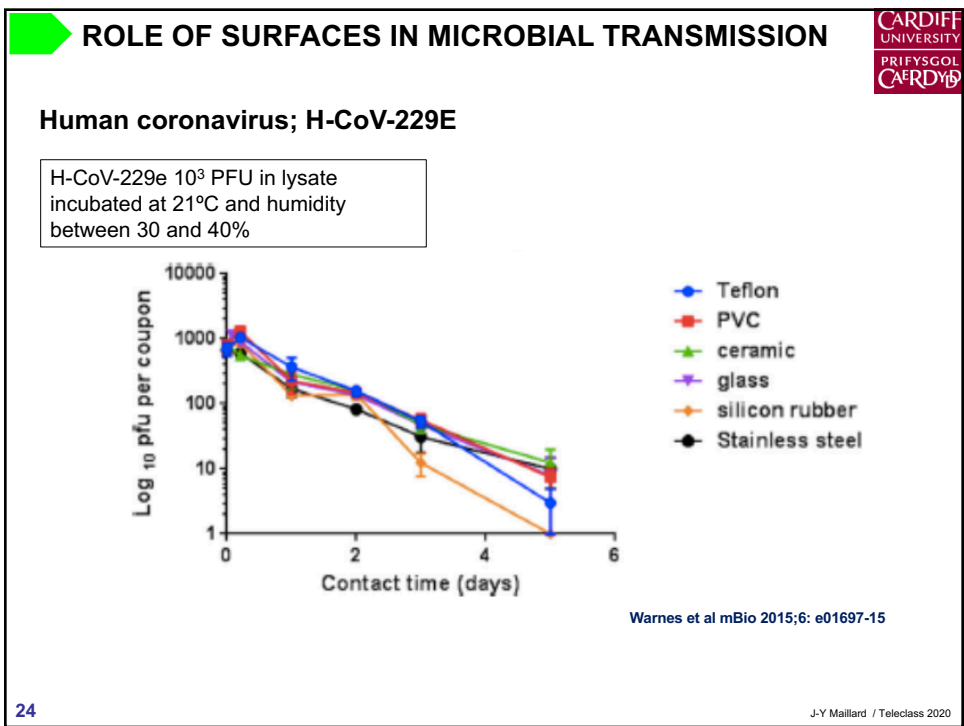
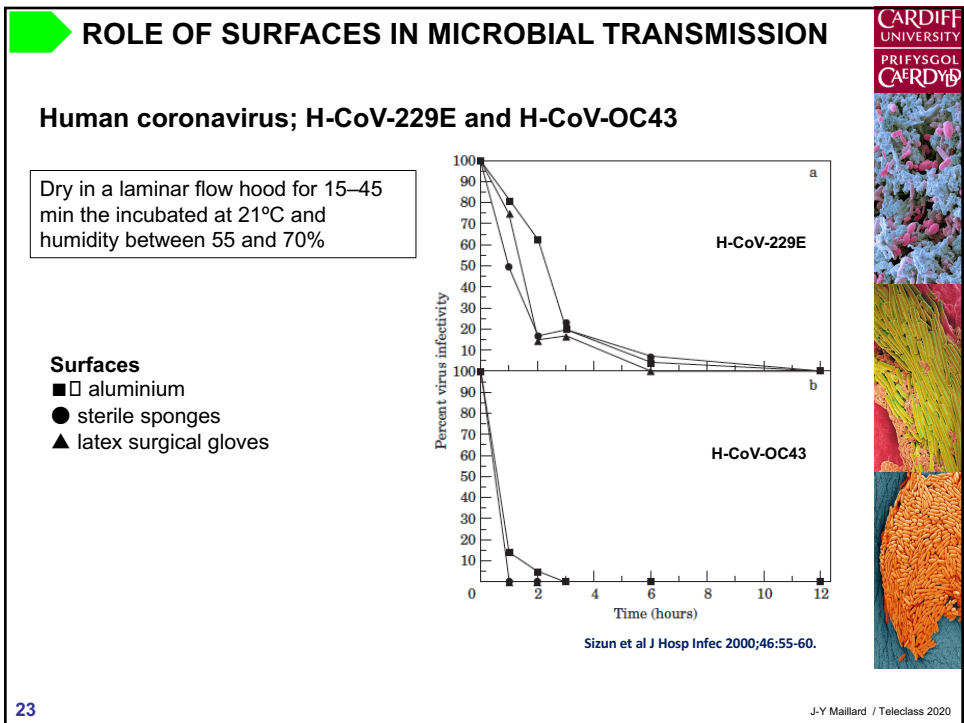
“Nevertheless, the agent’s (SARS-CoV) tenacity is considerably higher than that of HCoV-229E, and should SARS re-emerge, increased efforts need to be devoted to questions of environmental hygiene.”



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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**Viruses differ greatly in size and morphology**

Diagram showing the approximate size and shapes of different viruses.  
(From Horne, 1978)

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**Levels of 'germicidal' action**  
Spaulding classification, 1972

<b>High-level</b>	<b>Prions</b>	Creutzfeld-Jakob disease
	<b>Bacterial spores</b>	<i>Bacillus</i> spp., <i>Geobacillus</i> spp., Clostridia
<b>Intermediate-level</b>	<b>Protozoal cysts and oocysts</b>	<i>Cryptosporidium</i> spp., <i>Acanthamoeba</i> spp., <i>Entamoeba</i> spp.
	<b>Mycobacteria</b>	<i>Mycobacterium tuberculosis</i> , <i>M. avium intracellulare</i>
	<b>Non-enveloped viruses</b>	Poliovirus, parvovirus, papilloma virus, norovirus
	<b>Fungi</b>	<i>Aspergillus</i> spp., <i>Candida</i> spp.
	<b>Vegetative Gram-negative</b>	<i>Pseudomonas</i> spp., <i>Klebsiella</i> spp.
<b>Low-level</b>	<b>Protozoa</b>	<i>Cryptosporidium</i> spp., <i>Acanthamoeba</i> spp.
	<b>Vegetative Gram-positive</b>	Staphylococci, streptococci
	<b>Enveloped viruses</b>	Hepatitis B virus, HIC, HSV, SARS

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**HIV**

Labels: Vif, Vpr, Nef and p7, Protease, Lipid Membrane, Nucleocapsid, Reverse Transcriptase, Integrase, Transmembrane Glycoprotein, gp41, gp120 Docking Glycoprotein, Capsid, Matrix, Viral RNA Genome.

Icosahedral symmetry      Helical symmetry

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**Biocide**

**Virus**

**CAPSID PERMEABILITY**  
Penetration of the biocides inside the capsid and destruction of nucleic acid

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**Biocide**

**Virus**

**SPECIFIC TARGET 'PROTEINS'**  
Alteration of virus-host cell specificity followed by loss of infectivity

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

**Biocide**

**Virus**

**VIRUS STRUCTURE**  
Destruction of the capsid and nucleic acid

Destruction of the capsid and release of an intact nucleic acid

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

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**VIRUS STRUCTURE AND SUSCEPTIBILITY**

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## VIRUS STRUCTURE AND SUSCEPTIBILITY

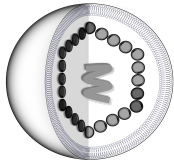


**ENVELOPE**

- Alkylating agents: interaction with viral proteins
- Membrane active agents (chlorhexidine, QAC, alcohols, surfactants, phenolics)


**STRUCTURAL ALTERATION**

- chlorine compounds (f2, rotavirus, poliovirus; HBV)
- bromine (reovirus type-3, f2, poliovirus)
- cetylpyridinium chloride (F116)
- cetrimide (rotavirus)
- alcohols (F116, rotavirus)
- phenols (F116)
- peracetic acid (F116, adenovirus)
- aldehydes (poliovirus, HBV)
- metallic salts (e.g. silver?)
- iodine (poliovirus, f2): tyrosine amino acid
- ozone (poliovirus type-2, ΦX174, T4, f2): cysteine, tryptophane, methionone (?)
- copper salts: thiol and other group of protein molecules
- GTA (F116, HBV, poliovirus): ε-amino acid of lysine residues




Maillard and Russell. *Sci Prog* 1997; 80: 287-315.  
 Maillard. *Rev Med Microbiol* 2001; 12: 63-74.

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## VIRUS STRUCTURE AND SUSCEPTIBILITY

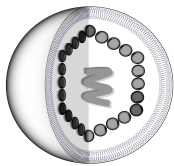


**VIRAL MARKERS**

- metallic salts: phage adsorption, DNA ejection
- alcohols: HbcAg, HBV DNA polymerase
- GTA: HbsAg & HbcAg, HBV DNA polymerase
- Chlorine compounds: HbsAg & HbcAg

**ALTERATION OF VIRAL GENOME**

- chlorine compounds (f2 RNA, poliovirus RNA)
- peracetic acid (F116 DNA)
- metallic salts (R17 RNA, ΦX174 DNA, poliovirus RNA)
- ozone (f2 RNA, ΦX174 DNA, T4 DNA): purines and pyrimidines
- GTA: HBV DNA (detection of HBV DNA residues)



Maillard and Russell. *Sci Prog* 1997; 80: 287-315.  
 Maillard. *Rev Med Microbiol* 2001; 12: 63-74.

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**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

**Definitions**

**Cleaning**

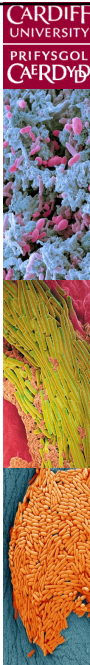
- Removal of contamination from a surface to the extent necessary for further processing or for intended use.

**Disinfection**

- Process of reduction of the number of viable microorganisms to a level previously specified
- Three levels of disinfection (high, intermediate and low)

**Sterilization**

- Validated process used to render a product/surface free from viable microorganisms, **including bacterial spores.**

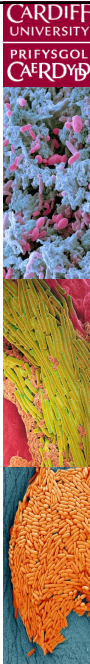


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**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

What level of efficacy is necessary?

High level of shedding Low infectious dose	High level of shedding High infectious dose
Low level of shedding Low infectious dose	Low level of shedding High infectious dose



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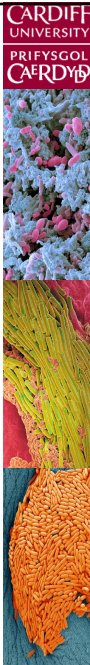
**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

What level of efficacy is necessary?

**High level of shedding**  
**Low infectious dose**

e.g. norovirus

Shedding:  $1 \times 10^8$  virus/ml or /g (8  $\log_{10}$ )  
ID:  $10^2$  virus (2  $\log_{10}$ )  
**>6  $\log_{10}$  reduction of 99.9999%**



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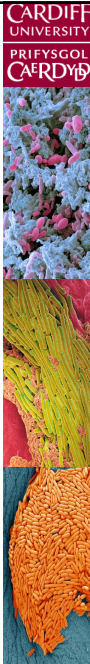
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**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

What level of efficacy is necessary?

Shedding:  $1 \times 10^8$  virus/ml or /g (8  $\log_{10}$ )  
ID:  $10^4$  virus (4  $\log_{10}$ )  
**>4  $\log_{10}$  reduction of 99.99%**

**High level of shedding**  
**High infectious dose**



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**How Difficult is it to Eliminate Coronavirus From Surfaces?**  
**Prof. Jean-Yves Maillard, Cardiff University, Wales**  
**A Webber Training Teleclass**

**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

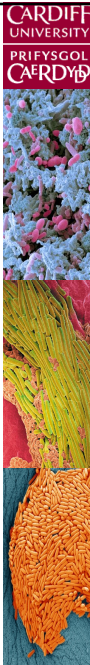
What level of efficacy is necessary?

Shedding:  $1 \times 10^5$  virus/ml or /g  
( $5 \log_{10}$ )

ID:  $10^4$  virus ( $4 \log_{10}$ )

> $1 \log_{10}$  reduction of 90%

Low level of shedding  
High infectious dose



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**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**

What level of efficacy is necessary?

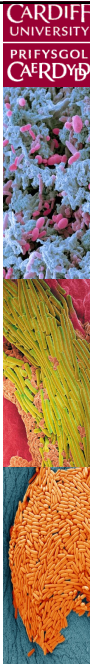
Shedding:  $1 \times 10^5$  virus/ml or /g  
( $5 \log_{10}$ )

ID:  $10^2$  virus ( $2 \log_{10}$ )

> $3 \log_{10}$  reduction of 99.9%

Low level of shedding  
Low infectious dose

e.g. SARS?



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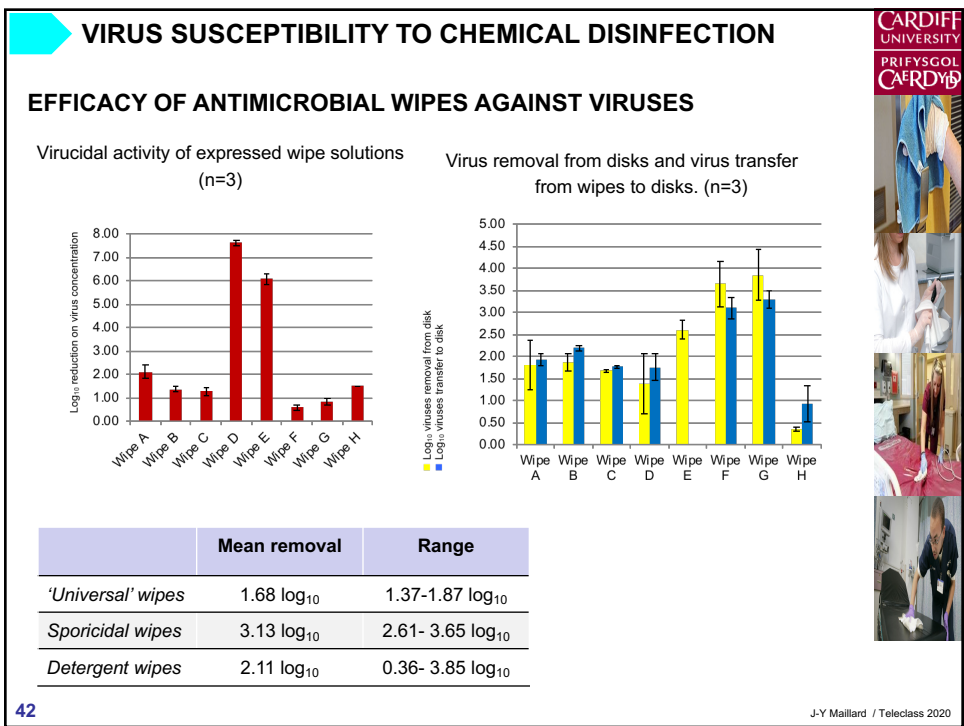
**VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION**



Disinfectant	Contact Time	Log <sub>10</sub> Reduction			
		Parvoviruses	Poliovirus	Adenovirus	Vaccinia
Alcohol (70%)	10 min	<1	2	>4	>4
QUAT (0.05%)	10 min	<1	<1	1	3
Bleach (1/10)	10 min	0.6 to 3	3	>4	>4
2% Glutaraldehyde	20 min	3 to 4	>4	>4	>4
0.55% OPA	10 min	3 to 4	>4	>4	>4
0.2% PAA (at 20°C)	10 min	>4	>4	>4	>4

Eterpi et al. J Hosp Infect 2009;73:64-70.

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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION

Number of wipes	Surface initially wiped	Time applied (sec)	Number of consecutive surfaces wiped (other surfaces)
1	Bed Rail	4	5 (bedside table, monitor X2, monitor stand)
2	Steel Trolley	6	2 (both shelves on the trolley wiped)
1	Monitor	4	5 (monitors, two keypads, monitor stand)
2	Bed rail	7	4 (table, monitor, keypad)
3	Bedside table	10	4 (folder, two bed rails)

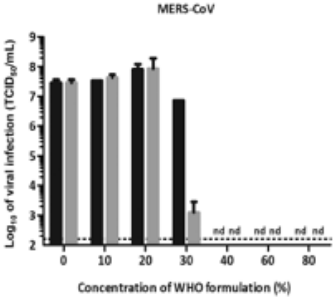
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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION

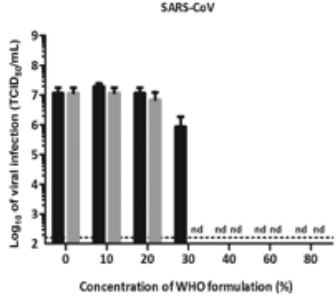
### SARS-CoV and MERS-CoV Suspension test

30 seconds contact time  
WHO ethanol-based formulations



MERS-CoV



SARS-CoV



Siddharta et al. J Infect Dis 2017;215:902-6

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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION






### Coronavirus SARS-CoV and PVP-I Suspension test

**Table 1.** Efficacy of PVP-I products to SARS-CoV

Reagent or treatment	Final PVP-I concentration, %	Virus titer after treatment, TCID <sub>50</sub> /ml	
		60 s	120 s
Control	0	1.17 × 10 <sup>6</sup>	n.d.
Isodine	1	95.1	u.d.
Isodine Gargle	0.47	190	u.d.
Isodine Scrub	1	u.d.	n.d.
Isodine Palm	0.25	u.d.	n.d.
Isodine Nodo Fresh	0.23	u.d.	n.d.
Ethanol (final 35%)	-	u.d.	n.d.

n.d. = Not done; u.d. = under the detectable level.

Kariwa et al. *Dermatology* 2006;212:119-23

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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION






### SARS-CoV Suspension test

**Table 2.** Viricidal activity of different disinfectants against SARS-CoV

Treatment	Virus titre (TCID <sub>50</sub> /ml [log <sub>10</sub> ]) (after contact time of x s)	Minimal reduction factor (log <sub>10</sub> )
2-Propanol <sup>a</sup> (100%)	≤ 1.8 ± 0 (30 s)	≥ 3.31
2-Propanol <sup>a</sup> (70%)	≤ 1.8 ± 0 (30 s)	≥ 3.31
Desderman <sup>b</sup> (78% ethanol)	≤ 1.8 ± 0 (30 s)	≥ 5.01
Sterillium <sup>c</sup> (45% 2-propanol, 30% 1-propanol)	≤ 3.8 ± 0 (30 s)	≥ 2.78
Wine vinegar <sup>d</sup>	≤ 2.80 ± 0 (60 s)	≥ 3.0
Formaldehyde (0.7%) <sup>b</sup>	≤ 3.8 ± 0 (120 s)	≥ 3.01
Formaldehyde (1.0%) <sup>b</sup>	≤ 3.8 ± 0 (120 s)	≥ 3.01
Glutaraldehyde (0.5%) <sup>b</sup>	≤ 2.8 ± 0 (120 s)	≥ 4.01
Incidin plus <sup>e</sup> (2%) (26% glucoprotamin)	≤ 4.8 ± 0 (120 s)	≥ 1.68

<sup>a</sup>Input virus titre 5.55 ± 0.44  
<sup>b</sup>Input virus titre 7.18 ± 0.37, tested by membrane filtration  
<sup>c</sup>Input virus titre 6.95 ± 0.37, tested by membrane filtration  
<sup>d</sup>Input virus titre 5.93 ± 0.13  
<sup>e</sup>Input virus titre 6.48 ± 0.37, tested by membrane filtration

Rabenau et al. *Med Microbiol Immunol* 2005;194:1-6.

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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION






### SARS-CoV

#### Suspension test

Rabenau et al. JHI 2005;61:107-11

Table I Efficacy of different types of disinfectant at various exposure times against SARS coronavirus, expressed as minimum reduction factors (RFs) of three parallel experiments: 0.3% serum albumin (BSA), 10% fetal calf serum (FCS), and 0.3% BSA with 0.3% sheep erythrocytes

Product	Type of area of application	Concentration	Exposure time	RF (and SD)		
				0.3% BSA	10% FCS	0.3% BSA and 0.3% sheep erythrocytes
Sterillium	Hand rub	Undiluted	30 s	≥4.25 (0.47)	≥4.25 (0.47)	≥4.25 (0.47)
Sterillium Rub	Hand rub	Undiluted	30 s	≥4.25 (0.47)	≥4.25 (0.47)	≥4.25 (0.47)
Sterillium Gel	Hand rub	Undiluted	30 s	≥5.5 (0.54)	≥5.5 (0.54)	≥5.5 (0.54)
Sterillium Virugard	Hand rub	Undiluted	30 s	≥5.5 (0.54)	≥5.5 (0.54)	≥5.5 (0.54)
Mikrobac forte	Surface disinfectant	0.5%	30 min	≥6.13 (0.35)	≥6.13 (0.35)	≥6.13 (0.35)
			60 min	≥6.13 (0.35)	≥6.13 (0.35)	≥6.13 (0.35)
Kohrsolin FF	Surface disinfectant	0.5%	30 min	≥3.75 (0.71)	≥3.75 (0.71)	≥3.75 (0.71)
			60 min	≥3.75 (0.71)	≥3.75 (0.71)	≥3.75 (0.71)
Dismozon pur	Surface disinfectant	0.5%	30 min	≥4.5 (0.54)	≥4.5 (0.54)	≥4.5 (0.54)
			60 min	≥4.5 (0.54)	≥4.5 (0.54)	≥4.5 (0.54)
Korsorex basic	Instrument disinfectant	4%	15 min	≥3.25 (0.47)	≥3.25 (0.47)	≥3.25 (0.47)
		3%	30 min	≥3.25 (0.47)	≥3.25 (0.47)	≥3.25 (0.47)
		2%	60 min	≥3.25 (0.47)	≥3.25 (0.47)	≥3.25 (0.47)

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## VIRUS SUSCEPTIBILITY TO CHEMICAL DISINFECTION

### ... Other studies


**Sattar et al. Epidem Inf 1989;102:498-505**

- H-CoV-229E compared to other viruses
- Virus on surfaces in mucin or faeces – 1 min exposure
- 3 log<sub>10</sub> reduction after 1 min exposure time ■ yes ■ no

NaOCl	0.10% (pH 9.4)
	0.5% (pH 11.0)
Chloramine T	0.10% (pH 8.0)
	0.30% (pH 8.0)
PVP-I	1% iodine (pH 3.0)
Ethanol	70%
GTA	2% (pH 7.0)

Chlorhexidine cetrimide	0.008%
Chlorhexidine cetrimide	0.08%
+ ethanol	70%

BZC	0.04% (pH 6.0)
BZC + HCl	0.04% (pH 1.0) 7%
BZC + ethanol	0.04% (pH 5.0) 70%
BZC + sodium metasilicate	0.04% (pH 11.0) 0.5%



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**CONCLUSIONS**

- Enveloped viruses are really susceptible to chemical disinfectant

**BUT**

Some membrane agents need to be formulated with other excipients to have rapid efficacy against SARS-CoV

- Enveloped viruses are generally fragile but can persist on surfaces when mixed with organic load



SARS-CoV may persist better at low temperature and low RH

- Required efficacy for a chemical disinfectant depends on:





**Virus type**  
**Virus load on surface**  
**Virus infectious dose**

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**THANK YOU**



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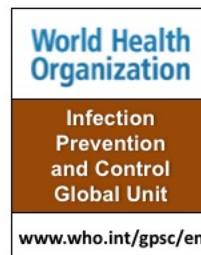
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<a href="http://www.webbertraining.com/schedule1.php">www.webbertraining.com/schedule1.php</a>	
June 17, 2020	<i>(South Pacific Teleclass)</i> <b><u>SHARPS INJURIES - WHY AREN'T WE AT ZERO?</u></b> Speaker: <b>Terry Grimmond</b> , Grimmond and Associates, New Zealand
July 9, 2020	<b><u>INFECTION PREVENTION AND CONTROL IN HOME CARE AND HOSPICE:            COMMON COMPLIANCE ISSUES</u></b> Speaker: <b>Mary McGoldrick</b> , Home Health Systems, Inc.
July 16, 2020	<i>(FREE Teleclass)</i> <b><u>THE BUZZ AROUND MOSQUITOES AND MOSQUITO-BORNE DISEASES</u></b> Speaker: <b>Dr. Marcia Anderson</b> , Environmental Protection Agency
July 23, 2020	<b><u>IMPROVEMENT OF HOSPITAL ENVIRONMENTAL CLEANING AND            DISINFECTION PRACTICES FOLLOWING AN EIGHT-MONTH OUTBREAK</u></b> Speaker: <b>Corey Weisgerber</b> and <b>Terrence Shaw</b> , Regina General Hospital, Saskatchewan
August 6, 2020	<b><u>CLEANING AND DISINFECTION IN THE ERA OF SARS-COV-2</u></b> Speaker: <b>Dr. Curtis Donskey</b> , Louis Stokes VA Medical Center, Cleveland, Ohio
August 13, 2020	<b><u>AHEAD - A CONSOLIDATED FRAMEWORK FOR BEHAVIOURAL INFECTIOUS            RISKS IN ACUTE CARE - PART 2</u></b> Speaker: <b>Prof. Hugo Sax</b> and <b>Dr. Lauren Clack</b> , University of Zurich Hospitals, Switzerland

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