

# Molecular Diagnostics and its Role in Infection Prevention

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A Webber Training Teleclass

## Molecular Diagnostics and its Role in Infection Prevention

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**NorthShore University Health System**

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April 6, 2018

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## The Changing Laboratory



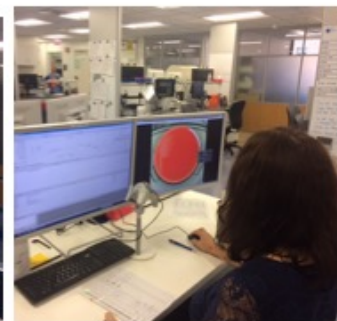
Late 1800's



Early 2000's



2013 →



Theparee et al. 2018. Total Laboratory Automation and MALDI-TOF Improve Turn-Around-Times in the Clinical Microbiology Laboratory: A Retrospective Analysis. J. Clin. Microbiol. 56:e01242-17.



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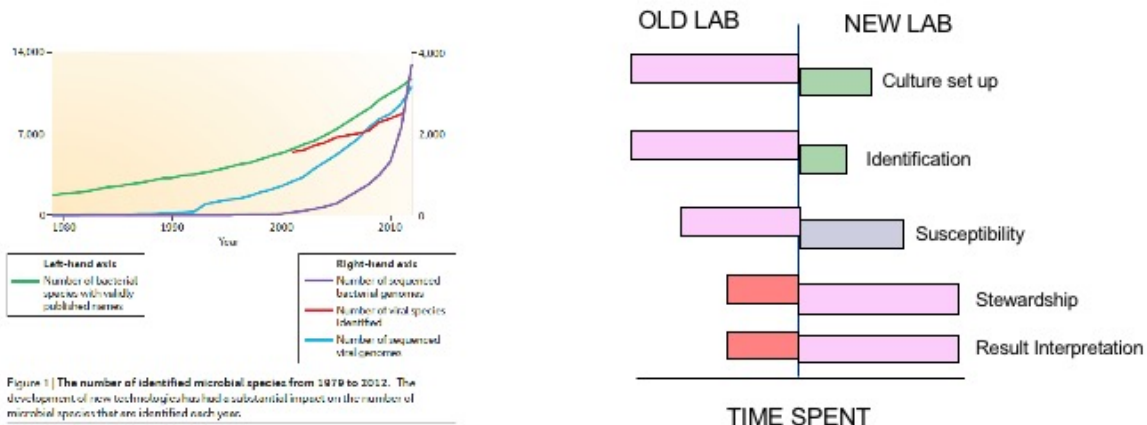
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## The Challenging Laboratory



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## Use of Molecular Assays in Infection Prevention

- Early diagnosis that can impact management
- Antimicrobial stewardship
- Screening and surveillance for Infection Prevention
- Choice of testing algorithms and platforms that improve infection control
- Implementation of new technologies (NGS): are we there yet?

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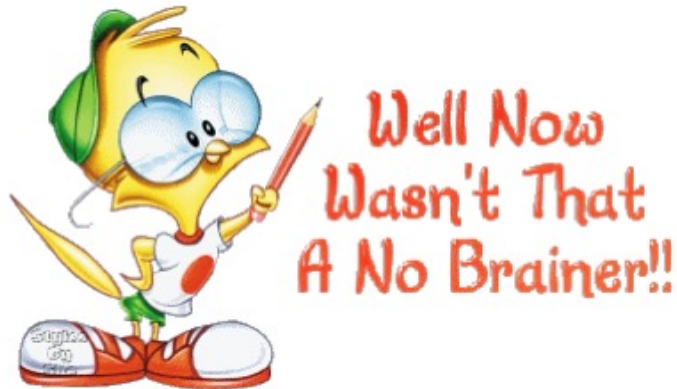
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## Early Diagnosis?

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Case

## Lung Abscess in a Young Adult

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- 37YO female former smoker presents on 11/17 with hemoptysis for 1 month
- Undergoing outpatient management of lung abscess
- Initially started on augmentin, later switched to clindamycin
- Bilateral chest pains worse with deep inspiration
- Rare coughing
- Generalized malaise, weakness and occasional nausea
- Underwent bronchoscopy on 10/27

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## Bronchoscopy Lab Results (10/27)

### Culture Results:

Streptococcus mitis group  
1+ rare  
Streptococcus salivarius group  
1+ rare  
Rothia mucilaginosa  
1+ rare

### Susceptibility Testing:

	R. mucilaginosa	S. mitis	S. salivarius
Ceftriaxone	S	S	R
Clindamycin	R	S	S
Erythromycin	S	R	R
Penicillin G	S	I	R
Tetracycline	S		
Vancomycin	S		
Trimeth/Sulfa	S	S	S

### Cytopathology

#### FINAL DIAGNOSIS:

Lung, right lower lobe (superior segment), transbronchial biopsy  
- Non-necrotizing granulomatous inflammation.  
- Acute and chronic inflammation.  
- No dysplasia or malignancy present.

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## Admission and Subsequent Management

- Admitted, started to vancomycin and piperacillin/tazobactam
- ID and Pulmonology consulted
- Chest Xray findings:
  - Large right perihilar mass similar in size to previous study. Left apical consolidation that may be due to pneumonia is smaller but still present. Heart and pulmonary vessels are normal in size. No pleural effusion.

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## Other Lab Results and Clinical Impression

- Fungal and vasculitis studies negative
- TB qunatiferon gold negative
- Non-TB mycobacteria possible
- Vancomycin intolerant
- Changed to linezolid and ertapenem

Lung abscess not responding to antibiotics  
Recommend surgery

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## VATS Laboratory Results (11/22)

### DIRECT SMEAR

Smear Result: Positive Acid fast bacilli 3+(10-90/field)

### HISTOPATHOLOGY

Right lower lobe of lung, wedge resection:  
- Necrotizing granulomatous inflammation. See comment and laboratory data.

#### COMMENT:

Sections taken from the right lower lobe of lung confirm the presence of the necrotizing granulomatous inflammatory process. An AFB stain is positive for numerous acid fast bacteria. Cultures were obtained from the lung parenchyma and AFB stains on that tissue are also positive. A Silver methenamine stain is negative for fungal elements.

TBPCR POSITIVE

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## Culture and Susceptibility (12/28)

### Culture Results:

M. tuberculosis complex

### Susceptibility Testing:

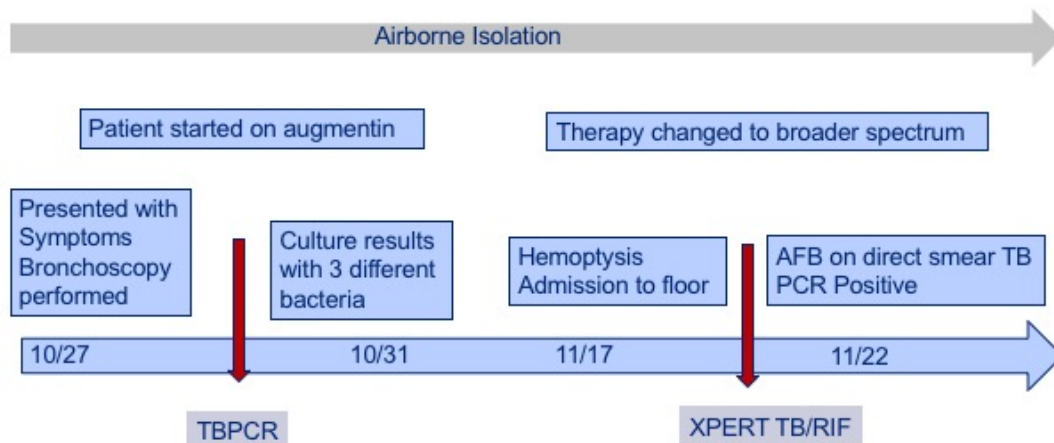
	M. tuberculosis
Capreomycin	S
Ciprofloxacin	S
Ethambutol	R
INH (L&H)	R
Kanamycin	S
Ofloxacin	S
Pyrazinamide	S
Rifampin	R
Streptomycin	R
Ethionamide	R

MDR-TB!  
Total time to Identification and Susceptibility  
~2 months

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## Timeline of Diagnosis and Management



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## WHO-RECOMMENDED DIAGNOSTIC TOOLS

**RECOMMENDED FOR USE** (detailed policy guidance: [http://www.who.int/tb/areas-of-work/laboratory/policy\\_statements/en](http://www.who.int/tb/areas-of-work/laboratory/policy_statements/en))

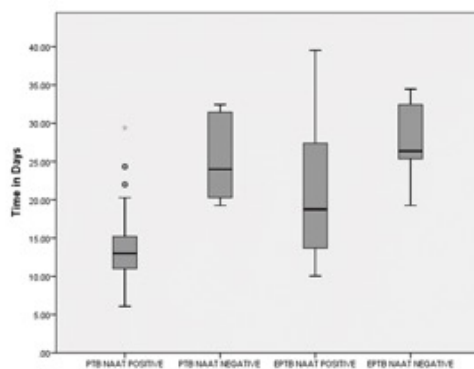
- **LED microscopy:** For use at all laboratory levels as replacement of conventional fluorochrome and light microscopy.
- **Commercial liquid culture and DST systems:** For use at central/regional reference laboratory level, as current reference standard.
- **Rapid speciation strip technology:** For use with conventional culture and DST at central/regional reference laboratory level, to identify *Mycobacterium tuberculosis*.
- **Automated real-time nucleic acid amplification - Xpert MTB/RIF system:** For rapid detection of pulmonary and extrapulmonary TB and rifampicin resistance in both adults and children at decentralised laboratory and health care centres.
- **Lateral flow urine lipoarabinomannan (LF-LAM) assay** may be used to assist in the diagnosis of TB in HIV positive patients with signs and symptoms of TB (pulmonary and/or extrapulmonary) who have a CD4 cell count less than or equal to 100 cells/ $\mu$ L, or HIV positive patients who are seriously ill regardless of CD4 count or with unknown CD4 count.
- **Loop-mediated isothermal amplification test kit for TB (TB-LAMP)** - manual molecular assay to replace microscopy to diagnose TB in settings where automated molecular tests cannot be used.
- **Line probe assay (LPA)** as a rapid diagnostic test for detection of rifampicin and isoniazid resistance. The WHO recommended commercially available tests include GenoType MTBDRplus VER 1 and 2 (Hain Lifescience, Germany), Nipro NTM+MDRTB detection kit 2 (Nipro, Japan). Suitable for use on smear-positive specimens or culture isolates.
- **Second-line line probe assay (SL-LPA)** as a rapid diagnostic test in patients with confirmed rifampicin-resistant TB or MDR-TB to detect resistance to fluoroquinolones and the second-line injectable drugs.

From: [www.who.int/tb](http://www.who.int/tb) Factsheet TB diagnosis

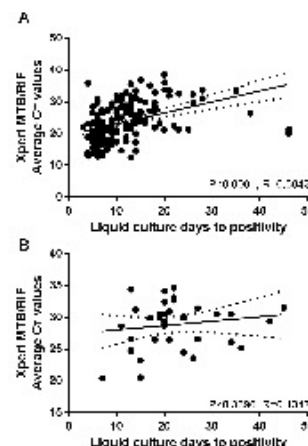


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## Advantages of Molecular Diagnosis: TB



Das, S. Abstract ID week 2017



Theron, G. Scientific Reports 2014



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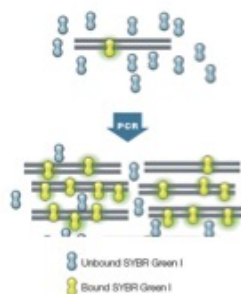
## Molecular Diagnosis of TB in Resource Limited Areas

- Advantages:
  - Rapidity of result delivery
  - Standardization of assay techniques
  - Potential for high throughput
  - Reduced requirements for biosafety
  - Sensitivity as high as 95% for some platforms
- Disadvantages:
  - Calibration
  - Instrumentation
  - Constant electric supply

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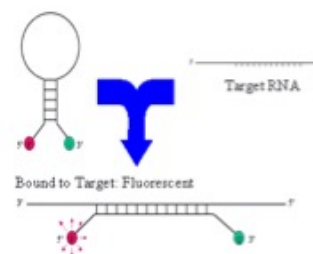
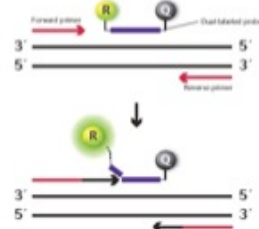
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## Real-Time PCR Assay Technologies: RECAP



### Advantages

- Minimizes cross-contamination
- Automated, low-complexity
- Quantitative
- High sensitivity



### Disadvantages

- Known pathogens only
- Only 2-6 plex
- Primer design: challenging for melt-curve
- Fluorescent-labeled primers: expensive

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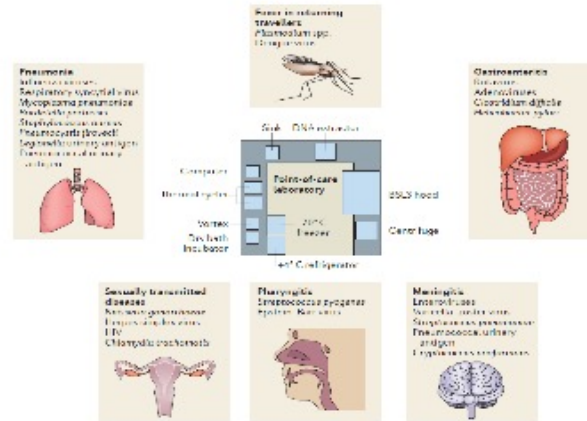
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## Syndromic Approach to Diagnostics



Fournier et al. Modern Clinical Microbiology Nature Reviews Microbiology 2013. 11:574.



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## The GI Panels

TABLE 1 Targets included on three commercial, FDA-cleared, multiplex assays for the detection of gastrointestinal pathogens

Target <sup>a</sup>	Multiplex panel <sup>b</sup>		
	Verigene EP	FilmArray GI	sTAG-GPP
<i>Aeromonas</i>		✓	✓
<i>Campylobacter</i>	✓	✓	✓
<i>Clostridium difficile</i> (toxin A/B)		✓	✓
<i>Plasmodium</i> spp.		✓	✓
<i>Salmonella</i>	✓	✓	✓
<i>Yersinia enterocolitica</i>	✓	✓	RIHO <sup>c</sup>
<i>Vibrio</i> spp.	✓	✓	✓
EPEC	✓	✓	✓
EPEC	✓	✓	✓
ETEC	✓	✓	✓
STEC (stx, and stx <sub>2</sub> )	✓	✓	✓
<i>E. coli</i> O157	✓	✓	✓
EPEC/Shigella	✓	✓	✓
<i>Cryptosporidium</i>	✓	✓	✓
<i>Cyclospora cayentensis</i>	✓	✓	✓
<i>Enterovirus histolytica</i>	✓	✓	✓
<i>Giardia lamblia</i>	✓	✓	✓
Adenovirus 40/41	✓	✓	✓
Norovirus GI/2GII	✓	✓	✓
Rotavirus A	✓	✓	✓
Sapovirus	✓	✓	✓
Astrovirus	✓	✓	✓

TABLE 2 Comparison of features between three commercial, FDA-cleared, multiplex platforms for the detection of gastrointestinal pathogens

Feature	Platform <sup>a</sup>		
	Verigene EP	FilmArray GI	sTAG-GPP
No. of FDA-cleared targets	9	22	14
Processing (hands-on) time per run, min	<5	2	45
Separate extraction required?	No	No	Yes (~45 min)
Time/run, h	~2	~1	~5
Throughput, specimens/run	1 <sup>b</sup>	1 <sup>b</sup>	96
Technology	PCR + gold nanoparticle hybridization	Nested PCR + melting curve	PCR + sTag (fluorescent bead-based detection)
Open or closed system	Closed	Closed	Open
Footprint	Small to moderate	Small	Moderate
List price per instrument, \$	40,000 <sup>c</sup>	39,500	37,000
List price reagent cost per specimen, \$	80 <sup>c</sup>	155 <sup>c</sup>	80-90 <sup>c</sup>

Binnicker et al. J. Clin. Microbiol. December 2015



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## The Problem with Diarrhea

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- *Issues with lab diagnosis*
  - Wide range of pathogens causing different types of diarrhea
  - Childhood diarrhea different from adult
  - A multitude of techniques needed for diagnosis
    - » ELISA for toxins
    - » Fluorescent microscopy for parasites
  - Global problem with varying etiology
- *Solution?*
  - Antimicrobial susceptibility and public health impact?
  - False positives with *Vibrio and Entamoeba*
  - Reimbursement: Do all patients need to get all 17 targets?
  - What will we miss if we move solely to panel based testing? (*Aeromonas*)

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## The Problem with Gold Standards: STI

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- PCR found to be 20-50% more sensitive for diagnosis of chlamydial infections than traditional culture methods
- Comparing results to an imperfect method produces biased sensitivity and specificity
- Prevalence being constant, sensitivity of older assays will be overestimated if standard tests are suboptimal
- Comparison with comparable technology
- Caveats?

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## Impact of Commensal Flora

Assay Name	Assay format	Stability of specimen at 2-30°C	Nucleic acid target	Comments
Abbott RealTime CT/NG (Abbott Laboratories, Abbott Park, IL)	Molecular beacon probe	14 days	Specific sequence within the C. trachomatis cryptic plasmid opa gene of N. gonorrhoeae.	Does not detect plasmid free C. trachomatis Commensal Neisseria species does not cause false positive reaction
Xpert CT/NG assay (Cepheid, Sunnyvale, CA)	Multiplexed Taqman assay	Variable depending on specimen type (8 hours to 45 days)		Detects plasmid free C. trachomatis Commensal Neisseria species does not cause false positive reaction
cobas CT/NG test (Roche Diagnostics, Indianapolis, IN)	Dual probe Taqman assay	Variable depending on specimen type (up to 1 year)	Two distinct genes one within the chromosome and other in the cryptic plasmid A highly conserved direct repeat region of N. gonorrhoeae called DRP-2	Detects plasmid free C. trachomatis Commensal Neisseria species does not cause false positive reaction*
Aptima COMBO 2 assay Aptima CT assay Aptima GC assay (Hologic/Diagn-Probe Inc., San Diego, CA)	Multiplexed Taqman assay	24 hours, 30-90 days if using transport cup	23S rRNA and 16S rRNA gene of C. trachomatis 16S rRNA gene of N. gonorrhoeae	Detects plasmid free and new variant C. trachomatis Commensal Neisseria species does not cause false positive reaction
BD ProbeTec ET CT/GC Amplified DNA assay (Becton Dickinson and Company, Sparks, MD)	Transcription-Mediated Amplification, and Dual Kinetic Assay	30 hours If urine transport cup used up to 90 days		Does not detect plasmid free C. trachomatis N. cinerea and N. lactamica might cause false-positive test results

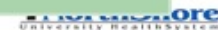
◆ Extra-genital *Chlamydia* and *Neisseria*: Impact the detection in pharyngeal rectal specimens impacts MSM, minors where detection in extra genital sites of importance



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## Respiratory Viral Panels

Assay name	Assay format/CLIA waived	Specimen	Extraction of nucleic acids required	Sensitivity/specificity	Pathogens detected
ProFAST® Assay (Prodesse®), Gen-Probe/Hologic)	Multiplex real-time reverse transcription PCR	NP swabs	Yes	94-99%	Depends on the panel used
FilmArray (BioFire)	Real-time PCR with melt curve analysis	NP swabs, respiratory secretions	No	84.5-100%	Adv: CoV HKU1, NL63; influenza virus A/B with typing; hMPV; PIV1, -2, -3, -4; RSV; RhV/EV; CoV HKU1 and NL63; Bordetella pertussis, Chlamydia pneumoniae, and Mycoplasma pneumoniae
Xpert flu (Cepheid)	Multiplex real-time reverse transcription PCR assay CLIA waived (Y)	NP swabs/Nasal aspirate	No	98-100%	Influenza A (H1), Influenza B
Simplera Flu A/B & RSV kit (Diasorin)	Multiplex real-time reverse transcription PCR	NP swabs	Yes	95%-99%	Influenza A/B and RSV
xTAGxTAG FAST™ (Luminex Corp.)	PCR, hybridization to fluorescent bead based liquid array	NP swabs	Yes	97-100%	Adv: influenza virus A (H1, H3); influenza virus B; MPV; PIV1, -2, -3; RSV (A/B); RhV/EV
eSensor® Respiratory Viral Panel (GenMark)	PCR followed by hybridization and electrochemical detection	NP swabs, respiratory secretions	Yes	98-99%	Influenza A, H1, 2009 H1, H3; Influenza B, Parainfluenza, RSV, human metapneumovirus, Rhinovirus and adenovirus
Vestigene RV Plus (Nanosphere)	Multiplex reverse transcription PCR followed by gold nanoparticle hybridization assay	NP swabs	No	96-100%	Influenza A, H1, 2009 H1, H3; Influenza B RSV,
Vestigene RV flex (Nanosphere)		NP swabs	No	NA	Influenza virus A/B with typing: hMPV; PIV1, -2, -3, -4; RSV; RhV; Adenovirus
Aleri (Abbott)	Isothermal PCR assay	NP and nasal swabs	No	88-100%	Influenza A/B
LIAT (ROCHE)	Multiplex real-time reverse transcription PCR assay CLIA waived (Y)	NP Swabs	No	98-100%	Influenza A/B and RSV



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## Elderly Gentleman with Pneumonia?

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- 85YO male in clinic on **13<sup>th</sup> January** with moderate productive cough for 1 day, fever up to a 100°F
- Chest X-ray ordered, patient prescribed **cefpodoxime** and advised to come back if worse
- Admitted on 14<sup>th</sup> of January
  - Fever of 101.5, congestion, generalized weakness
  - Concern for pneumonia
  - Antibiotics: **vancomycin** and **zosyn** continued
  - Blood culture and NP swab for respiratory virus were sent



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## Acute Exacerbation of Asthma?

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- 65YO female with asthmatic bronchitis presents to ED on **25<sup>th</sup> January** with complaints of 5 days of cough and shortness of breath
- Associated wheezing, nausea, vomiting, diarrhea and body ache
- Started on nebulizer and IV steroids-- minimal improvement; transferred to ICU for further management
- NP swab sent for respiratory viruses



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## Fever in an Infant

- Otherwise healthy 12 month old female presents *to Urgent Care Center* on **February 6<sup>th</sup>** with fever since waking this morning
- No other symptoms
- Does not attend day care, no sick contacts at home
- Vaccinations up-to-date
- NP swab sent for respiratory viruses

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## Laboratory Testing Viruses: The Past



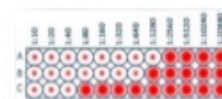
Specimen



Chorioallantoic Membrane & Allantoic cavity



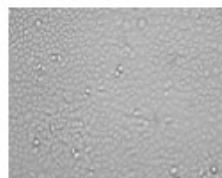
Harvest from infected egg  
~1 week



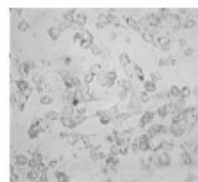
Hemagglutination/Hemagglutination inhibition



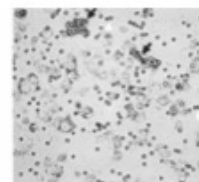
Viral Cell Culture Flask



Uninfected Cell Line



Viral Cytopathic Effect  
(~1 week)



Hemadsorption to RBC for identification

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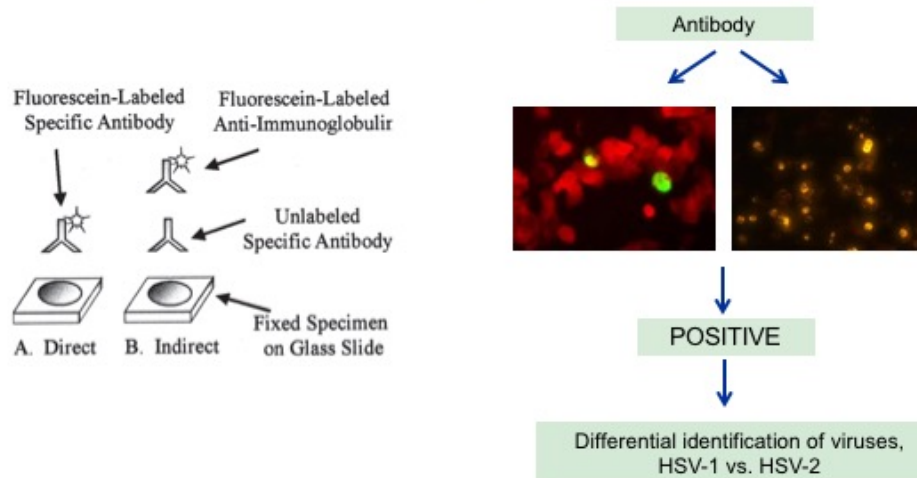
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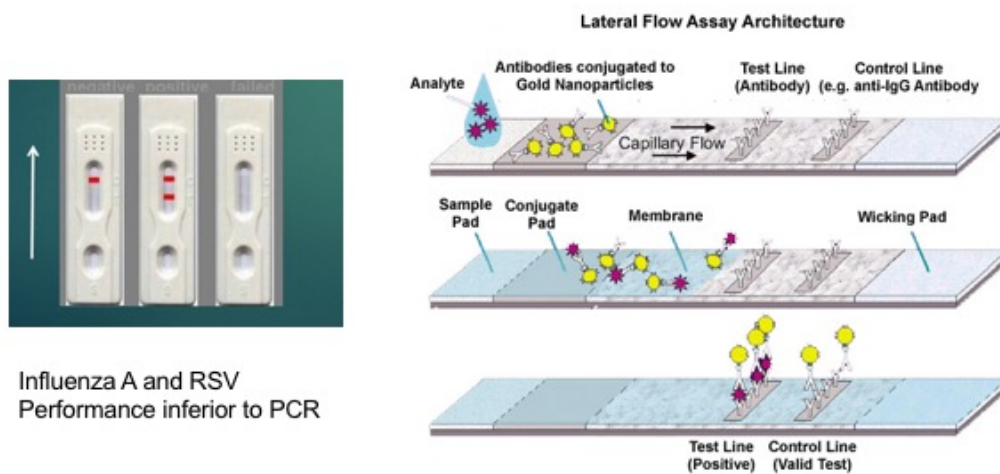
## Direct Fluorescent Antibody



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## Rapid Antigen Immunoassay



Influenza A and RSV  
Performance inferior to PCR

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## Laboratory Testing Viruses: Present


 	 	 
6-24 hours	3-4 hours	20-60 mins

**Turn around time**



30

<i>Complex</i>	<i>Simple</i>	
 	 	 
6-24 hours	3-4 hours	20-60 mins
<i>Centralized</i>	<i>Point-of care</i>	
Early 2000's	2014	2016-2017



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## Our Patients' Lab Results

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### Case 2

Elderly inpatient with CAP

Positive for Influenza A by PCR  
Negative for Influenza B by PCR  
Negative for RSV by PCR

### Case 3


Asthmatic in ICU

Influenza A	Positive
Influenza A H1	Not Detected
Influenza A H3	Positive
Influenza B	Not Detected
Respiratory Syncytial virus (RSV) A	Not Detected
Respiratory Syncytial virus (RSV) B	Not Detected
Parainfluenza (PIV) 1	Not Detected
Parainfluenza (PIV) 2	Not Detected
Parainfluenza (PIV) 3	Not Detected
Parainfluenza (PIV) 4	Not Detected
Human Metapneumovirus (hMPV)	Not Detected
Human Rhinovirus (HRV)	Not Detected
Adenovirus	Not Detected

### Case 4

Infant in ACC

Positive Influenza A detected  
Negative Influenza B not detected



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## Our Patients' Lab Results

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### Case 2

Elderly inpatient with CAP

**Turn around time up to ~6h**  
**\$**

Positive for Influenza A by PCR  
Negative for Influenza B by PCR  
Negative for RSV by PCR

### Case 3

Asthmatic in ICU

**Turn around time ~3-4h**  
**\$\$\$**


Influenza A	Positive
Influenza A H1	Not Detected
Influenza A H3	Positive
Influenza B	Not Detected
Respiratory Syncytial virus (RSV) A	Not Detected
Respiratory Syncytial virus (RSV) B	Not Detected
Parainfluenza (PIV) 1	Not Detected
Parainfluenza (PIV) 2	Not Detected
Parainfluenza (PIV) 3	Not Detected
Parainfluenza (PIV) 4	Not Detected
Human Metapneumovirus (hMPV)	Not Detected
Human Rhinovirus (HRV)	Not Detected
Adenovirus	Not Detected

### Case 4

Infant in ACC

**Turn around time <1h**  
**\$\$**

Positive Influenza A detected  
Negative Influenza B not detected





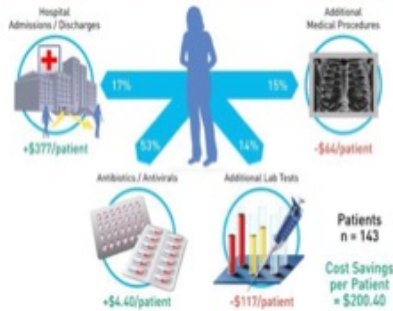
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## Impact on Clinical Decision

### Influenza Testing in the Emergency Department: Four Critical Touch Points



**Table 3**  
Impact of Rapid Influenza testing on clinical decision making in the ED for suspected Influenza Patients (n = 143)

Clinical Touchpoint	Total cases impacted (%) n/N* (N=143)	% reduction in utilization/change in discharge	% increase in utilization or admission
Antimicrobial prescribing total	26% (33/143)	10% (3/143) ↓	15% (21/143) ↑
Antibiotic prescribing	38.9% (56/143)	9% (13/143) ↓	88.6% (48/143) ↑
Antiviral prescribing	20.2% (29/143)	24.5% (33/143) ↓	14.7% (21/143) ↑
Medical Procedures/Imaging	15.4% (22/143)	23% (2/143) ↓	13.2% (19/143) ↑
Laboratory studies	14% (20/143)	3.8% (1/143) ↓	11.1% (16/143) ↑
Hospital Admissions/Discharge	18% (26/143)	10.5% (13/143) ↓	7.7% (11/143) ↓

Based on total number of ED patients meeting all inclusion criteria (N = 143).  
\*P < .05, binomial one sample test where the null hypothesis was Proportion = 0.  
†Antimicrobial prescribing refers to rates of antibiotic and antiviral prescribing.

Hansen GT, et al. 2018. Clinical decision making in the emergency department setting using rapid PCR: Results of the CLADE study group. J. Clin Virology; 102:42-49



34

## Impact on Clinical Decision: Outpatient Study

Test Result	Number	Prescribed Medications				p-value
		Antiviral*	Antibiotic**	None	Both	
<b>LIAT (POC-PCR) – Site A</b>						
Negative	128	3	57	68	0	*<0.0001
Positive (FluA)	63	47	2	9	5	
Positive (FluB)	51	38	3	6	9	
<b>RADT (POC-Antigen) – Site B</b>						
Negative	108	32	38	48	10	
Positive (FluA)	82	69	3	12	2	
Positive (FluB)	51	33	6	16	4	

Significant difference in antiviral prescription for patients with negative LIAT

Benirschke et al.; Abstract: Clinical Virology Symposium 2018



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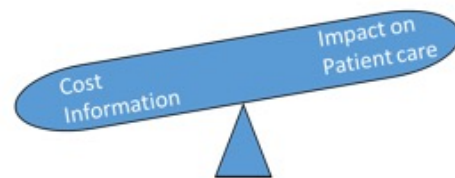
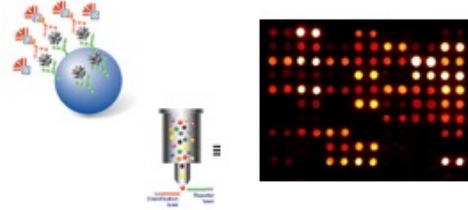
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## Multiplexed PCR Assays: Syndromic approach

- Concept of one test for one diagnosis:
  - Pneumonia panel
  - Sepsis panel
  - Diarrhea panel
  - Meningitis panel
- Advanced technology:
  - Appeal of complete automated sample handling
  - Easy read-out and software for analysis
- Strategic combination of targets:
  - Short time for broad range of pathogens
  - Provides some anti-infective information



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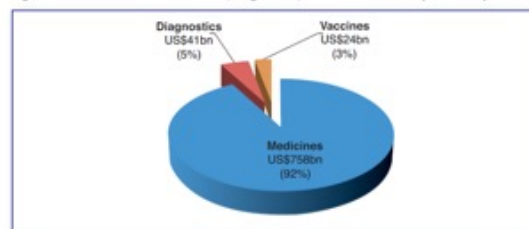
## Antimicrobial Stewardship: A lost Cause?



The Economist, March 31<sup>st</sup>, 2011

*"There is probably no chemotherapeutic drug to which in suitable circumstances the bacteria cannot react in some way acquiring 'fastness' [resistance]"* ---- Alexander Fleming, 1946

Figure 6.2 Global markets of medicines, diagnostics, and vaccines in 2006 (US\$ billion)



WHO, April 2014

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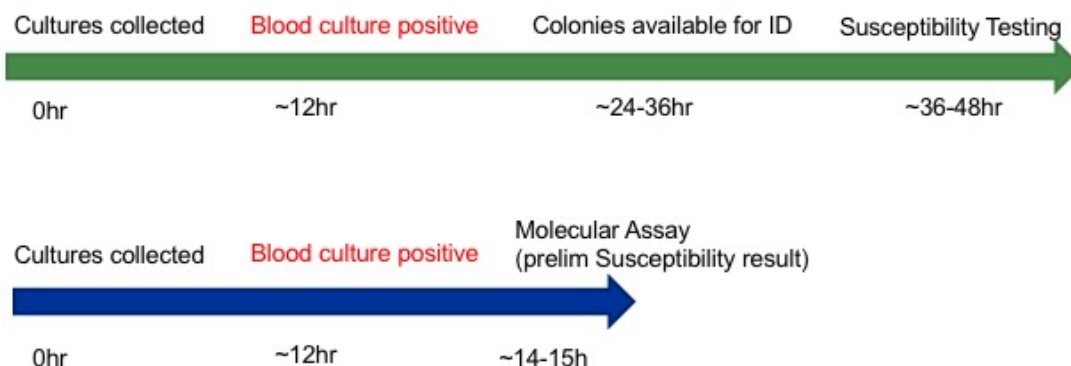
## Elderly Gentleman with Sepsis

- 76 year old male from nursing home, Parkinson's disease
- On chronic Foley catheter, and failure to thrive
- Brought to ED for fever, low blood pressure
- Alert and oriented but hypotensive
- Laboratory Results
  - Serum lactate level: 6.3
  - WBC count: 30,000/ul
- Working Diagnosis: Sepsis, blood cultures sent and started on 3<sup>rd</sup> generation cephalosporin

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## Use of Molecular Assays on Blood Culture



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## Molecular Assays for Antimicrobial Stewardship

- Elderly sepsis patient
  - E. coli by molecular assay (14h after blood culture collection)
  - Prelim result CTXM positive
  - 3<sup>rd</sup> generation cephalosporin changed to carbapenem
    - » De-escalated to beta-lactamase inhibitor combination after full susceptibility

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## Clinical Decision Support

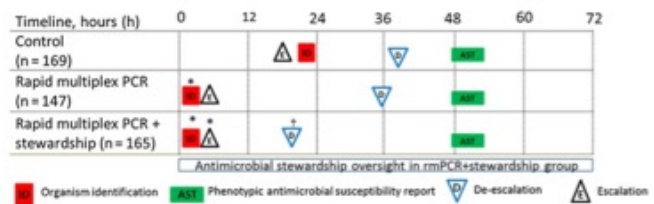
### Molecular Assay vs. Conventional Susceptibility:

Negative Predictive Values for ceftriaxone susceptibility in *Escherichia coli* and *Klebsiella pneumoniae* in the absence of either CTX-M or a carbapenemase gene were 98% and 93 to 94%, respectively.

Similar results were seen with other target bug-drug scenarios, with NPVs of 94 to 100% with the exception of *P. aeruginosa*, for which NPVs were poor, likely due to the more complex nature of resistance in this pathogen.

Pogue JM et al. Antimicrob Agents Chemother. 2018 Apr 26;62(5)

### Molecular Assay vs. Conventional Susceptibility



Banerjee R, et al. Clin Infect Dis. 2015 Oct 1; 61(7): 1071–1080.

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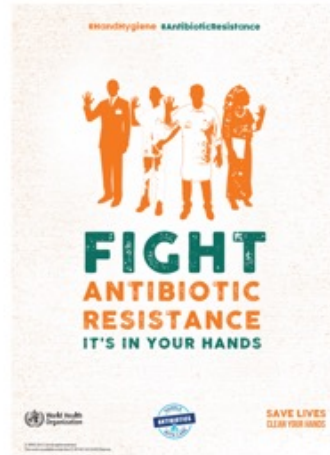
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## Infection Control Efforts and Surveillance



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## MRSA: The Success Story

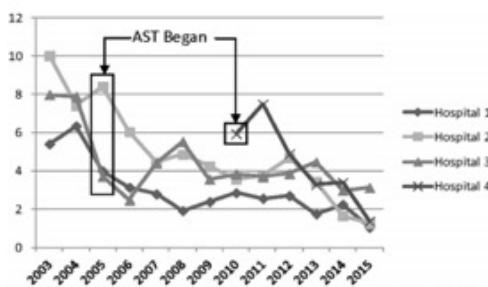


FIG 1 Rate of MRSA nosocomial infection (per 10,000 patient days). When calculated from baseline for each hospital, the aggregate  $P$  value was  $<0.001$ .

Peterson L, J. Clin. Microbiol. November 2016 Volume 54

### Impact of active screening and decolonization in ICU

- MRSA infection decreased from 3.58 to 0.42% ( $p < 0.05$ )
- Interruption of active surveillance and decolonization led to resurgence of infection rates up to 2.21%
- Reintroduction of intervention reduced in-hospital MRSA infection rates to 0.18%
- Decolonization and active surveillance saved \$22 for each \$1 spent on the intervention

Lee et al. Critical Care (2015) 19:143

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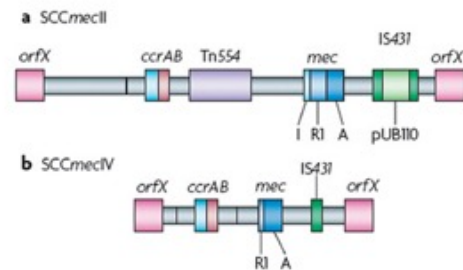
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## MRSA: The Success Story

- What qualities ensure success of MRSA screening test?
  - PCR test with quick result makes intervention simple and impact significant
  - Nasal swabs simple to collect and easy to test (not messy)
  - Site specific insertion of resistance gene, easy PCR design
  - Buy in from hospital and public health authorities



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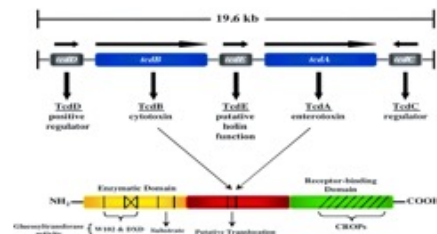
44

## C. difficile: The failure

 **HHS Public Access**  
Author manuscript  
*JAMA Intern Med.* Author manuscript; available in PMC 2016 July 18.  
Published in final edited form as:  
*JAMA Intern Med.* 2015 November ; 175(11): 1792–1801. doi:10.1001/jamainternmed.2015.4114.

### Overdiagnosis of *Clostridium difficile* Infection in the Molecular Test Era

Christopher R. Polage, MD, MAS, Clare E. Gyorke, BS, Michael A. Kennedy, BS, Jhansi L. Leslie, BS, David L. Chin, PhD, Susan Wang, BS, Hien H. Nguyen, MD, MAS, Bin Huang, MD, PhD, Yi-Wei Tang, MD, PhD, Lenora W. Lee, MD, Kyoungmi Kim, PhD, Sandra Taylor,



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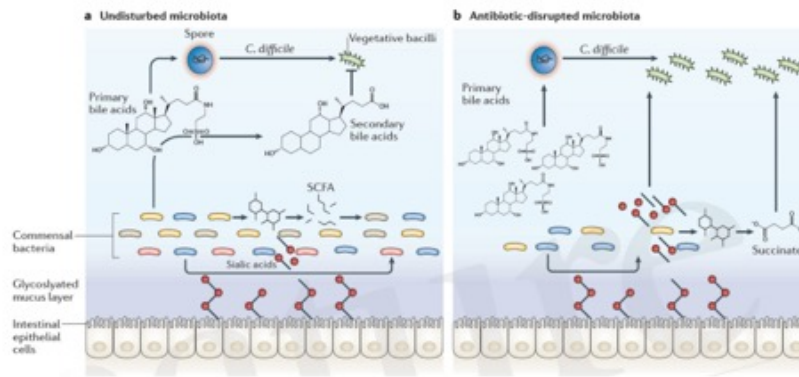
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## The difference is in the disease!



Abt MC, et al. *Nature reviews Microbiology*. 2016;14(10):609-620. doi:10.1038/nrmicro.2016.108.

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## The Art of Assay Design: *Unlimited?*

- Ideal Molecular Assay: Key Challenges
  - Rapid with relatively low technical complexity
  - **Minimal handling of samples**
  - **High throughput low turnaround time**
  - **A strategic combination of molecular targets based on specimen type**
  - Multiplexing: a panel for a syndrome
  - Guide to anti-infective therapy
  - Cost effective
  - Easy readout, software for analysis

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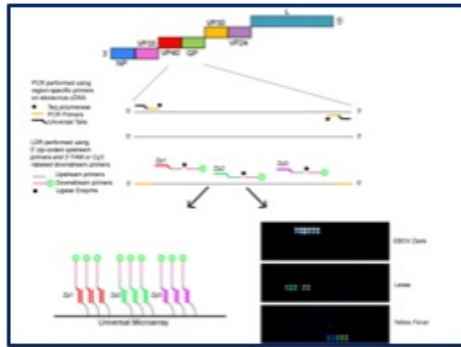
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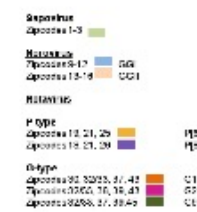
47

## The Art of Assay Design: *Unlimited?*



Das, S. et al. A Multiplex PCR/LDR Assay for the Simultaneous Identification of Category A Infectious Pathogens: Agents of Viral Hemorrhagic Fever and Variola Virus. *PLoS One*, 10(9):e0138484

Sample ID	1	2	3	4	5	6
1	10.00	1.50	1.50	0.00	0.00	0.00
2	11.11	1.11	4.00	1.41	0.01	1.11
3	6.92	1.50	1.50	1.4	0.04	1.4
9	0.01	0.09	0.04	1.00	0.03	0.00
10	0.01	0.06	0.01	1.14	0.04	1.14
11	1.11	1.25	0.01	1.36	1.03	1.00
12	1.14	0.01	0.01	1.40	0.01	1.40
14	0.19	0.20	1.4	1.4	0.01	1.4
15	0.01	0.01	0.11	0.01	0.01	0.01
16	0.00	0.00	1.00	1.00	0.01	1.00
18	0.01	0.01	0.01	0.01	0.01	0.01
21	0.01	0.01	0.01	0.01	0.01	0.01
22	0.01	0.01	0.01	0.01	0.01	0.01
23	0.01	0.01	0.01	0.01	0.01	0.01
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26	0.01	0.01	0.01	0.01	0.01	0.01
27	0.01	0.01	0.01	0.01	0.01	0.01
28	0.01	0.01	0.01	0.01	0.01	0.01
29	0.01	0.01	0.01	0.01	0.01	0.01
30	0.01	0.01	0.01	0.01	0.01	0.01
31	0.01	0.01	0.01	0.01	0.01	0.01
32	0.01	0.01	0.01	0.01	0.01	0.01
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41	0.01	0.01	0.01	0.01	0.01	0.01
42	0.01	0.01	0.01	0.01	0.01	0.01
43	0.01	0.01	0.01	0.01	0.01	0.01
44	0.01	0.01	0.01	0.01	0.01	0.01
45	0.01	0.01	0.01	0.01	0.01	0.01
46	0.01	0.01	0.01	0.01	0.01	0.01
47	0.01	0.01	0.01	0.01	0.01	0.01
48	0.01	0.01	0.01	0.01	0.01	0.01
49	0.01	0.01	0.01	0.01	0.01	0.01
50	0.01	0.01	0.01	0.01	0.01	0.01



Mirza A., Das, S. et al. A Multiplex PCR/LDR Assay for Viral Agents of Diarrhea with the Capacity to Genotype Rotavirus. *In Press, Scientific Reports (Nature)*, Accepted July 2018



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## Fever Spikes....Again?!!!

### Case

<b>Present Illness</b> <p>48y/o female presents with pyelonephritis Nephrostomy tubes placed Drained fluid sent for culture Cultures negative Sent home on p/o antibiotics</p>	<b>Disease Course</b> <p>Comes back to ED in a few hours for fever and pain High WBC count 16000 Fever up to a 103 Blood, urine and drain fluid sent for culture Rocephin and Vanco started All cultures negative</p>	<b>Management</b> <p>Continues to be febrile to 102.6 Pain at site of nephrostomy and back Nephrostomy tube draining well WBC 15.2 CRP 256</p>
--	---	--

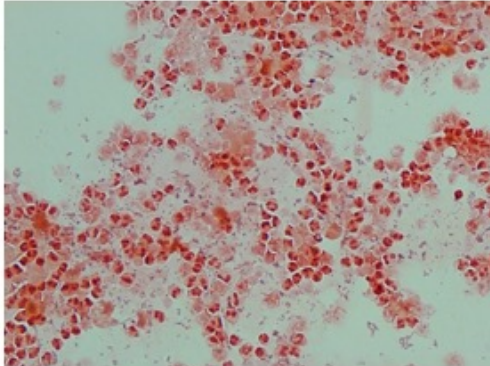




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## Gram Stain and Culture of Nephrostomy Drainage



4+ WBCs (PMNs)  
 No organisms seen

Culture Results:  
 Aerobic and anaerobic culture  
 No organism isolated

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## The Panbacterial Target: 16SrRNA Gene



Culture Results:  
 Aerobic and anaerobic culture  
 No organism isolated  
 Ureaplasma species. Isolate Comment: Detected by 16S rRNA gene sequencing. Results discussed with ID physician

Description	Max score	Total score	Query cover	E value	Ident	Accession
Ureaplasma urealyticum serovar 8 strain ATCC 27618 16S ribosomal RNA gene, partial sequence	898	898	98%	0.0	99%	<a href="#">NR_041718.1</a>
Ureaplasma parvum strain ATCC 27815 16S ribosomal RNA, complete sequence	863	863	98%	0.0	98%	<a href="#">NR_074762.2</a>

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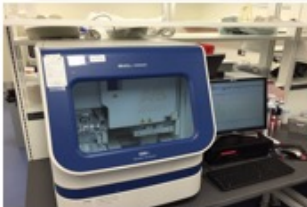
## The Future: New Technology?



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## Next Generation Sequencing: The Future?



Genetic analyzer: Sanger Sequencing



Palm top whole genome sequencer

- Ribosomal RNA gene sequence is used for ID
- Sequence information of whole genome
- Known pathogens and novel organisms
- Identify mutations for virulence and resistance
- Trace spread of pathogens

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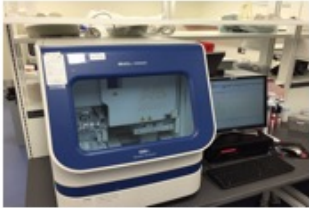
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## Next Generation Sequencing: The Future?



Genetic analyzer: Sanger Sequencing



Palm top whole genome sequencer

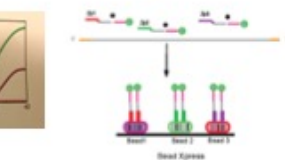
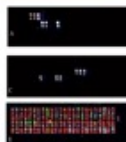
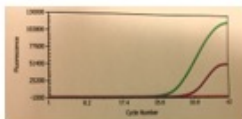
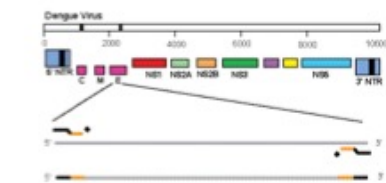
- Ribosomal RNA gene sequence is used for ID
- Sequence information of whole genome
- Known pathogens and novel organisms
- Identify mutations for virulence and resistance
- Trace spread of pathogens

- Reduction in running cost
- Clinical interpretation of data
- Streamlining bioinformatics and data analysis

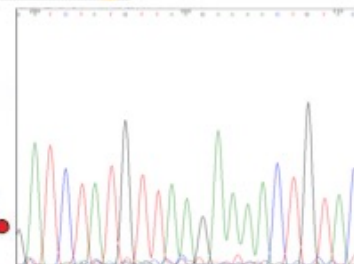
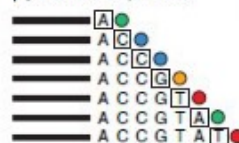
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## Targeted PCR vs. Sanger Sequencing



Dye terminator products



Identification in few hours

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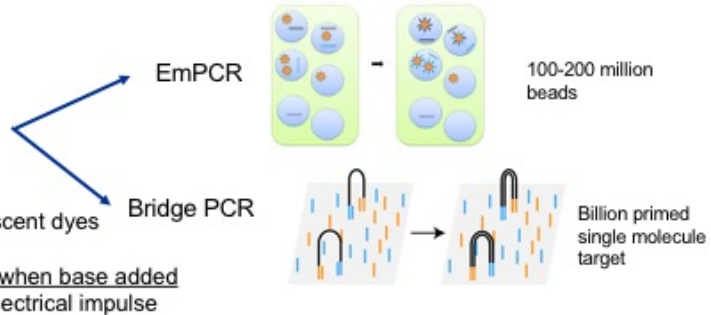
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## Next Generation Sequencing: Workflow

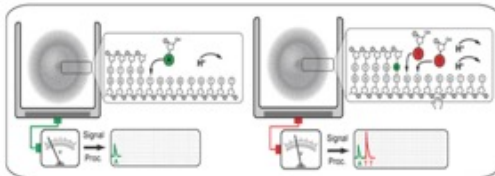
- Template Preparation
  - Randomly break into fragments
  - Templates need to be amplified
- Sequencing and Imaging
  - Reversible termination with fluorescent dyes
  - Ligation with fluorescent dyes
  - Semiconductor that measures pH when base added
  - Nanopore measures changes in electrical impulse
- Data Analysis
  - Align sequence to a reference database OR assemble *de novo*



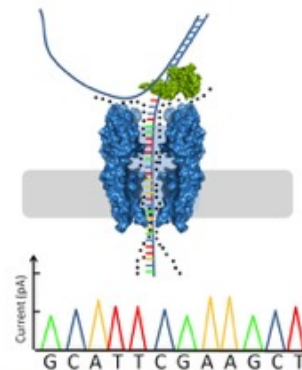
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## Next Generation Sequencing: Workflow



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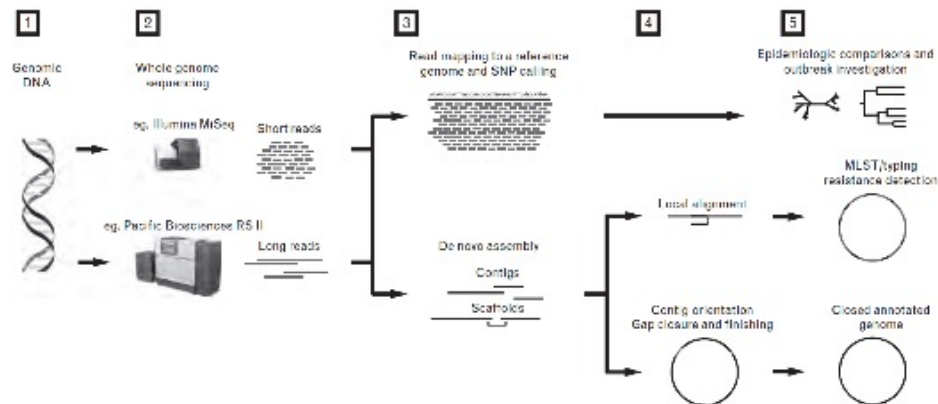
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## Next Generation Sequencing: WGS



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## Genotype: Phenotype Correlation Simplified?

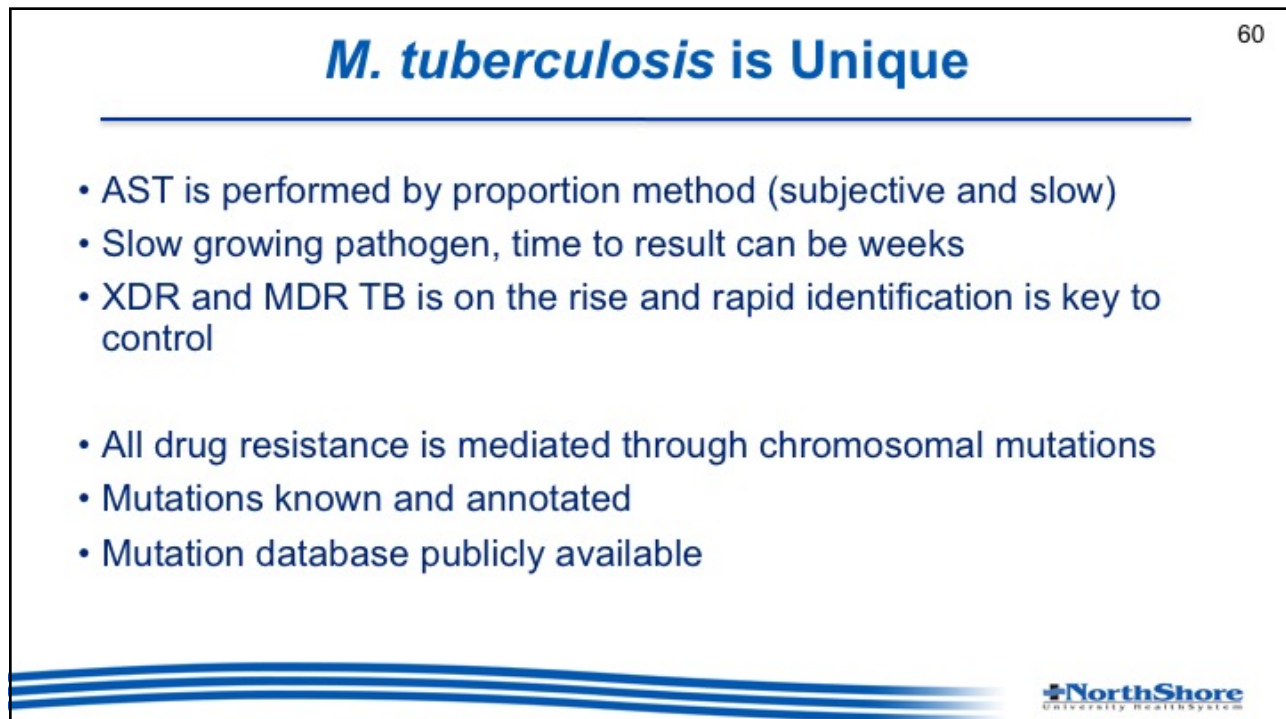
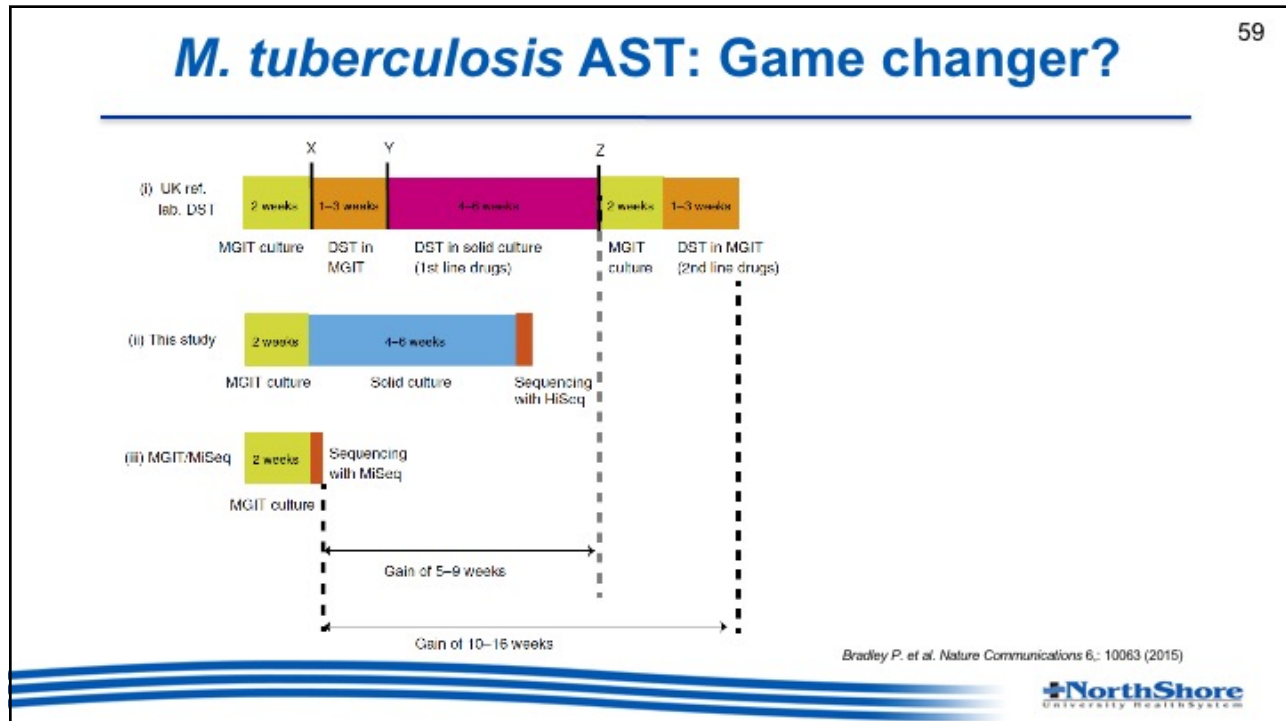
- How often does genotype predict resistance clinically?
- Can population genome graphs be used for identification and susceptibility?
- Is it better or rapid compared to existing methods?
- Algorithmic approach could work for some pathogens?

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Review

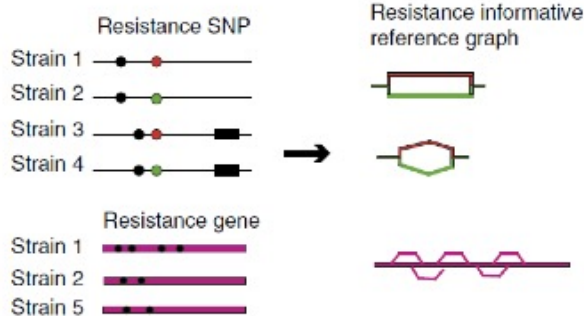
The role of whole genome sequencing in antimicrobial susceptibility testing of bacteria: report from the EUCAST Subcommittee

M.J. Ellington<sup>1,†</sup>, O. Ekelund<sup>2,†</sup>, F.M. Aarestrup<sup>3</sup>, R. Canton<sup>4</sup>, M. Doumith<sup>1</sup>, C. Giske<sup>5</sup>, H. Grundman<sup>6</sup>, H. Hasman<sup>7</sup>, M.T.G. Holden<sup>8</sup>, K.L. Hopkins<sup>1</sup>, J. Iredell<sup>9</sup>, G. Kahlmeter<sup>2</sup>, C.U. Köser<sup>10</sup>, A. MacGowan<sup>11</sup>, D. Mevius<sup>12,13</sup>, M. Mulvey<sup>14</sup>, T. Naas<sup>15</sup>, T. Peto<sup>16</sup>, J.-M. Rolain<sup>17</sup>, Ø. Samuelsen<sup>18</sup>, N. Woodford<sup>1,\*</sup>

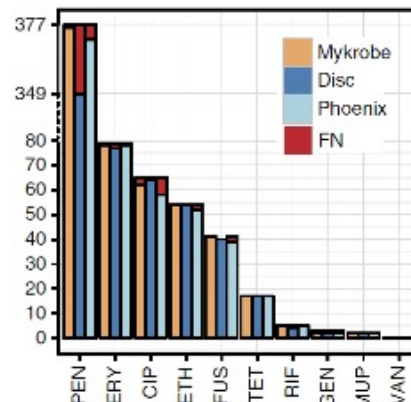
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## Rapid Antibiotic Resistance Predictions

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Build a graph of loci of interest  
Mark R loci (red) and S loci (green), predict by statistical algorithm



AST Prediction for *S. aureus* by Mykrobe

Bradley P. et al. Nature Communications 6: 10063 (2015)

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## Single nucleotide changes do not reflect plasmid mediated resistance

*Sci Transl Med* 2014 September 17; 6(254): 254ra126. doi:10.1126/scitranslmed.3009845

### Single molecule sequencing to track plasmid diversity of hospital-associated carbapenemase-producing Enterobacteriaceae

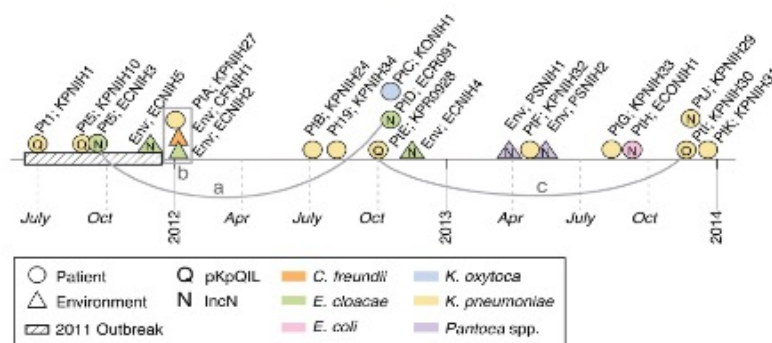
Sean Conlan<sup>1</sup>, Pamela J. Thomas<sup>2</sup>, Clayton Deming<sup>1</sup>, Morgan Park<sup>2</sup>, Anna F. Lau<sup>3</sup>, John P. Dekker<sup>3</sup>, Evan S. Snitkin<sup>1</sup>, Tyson A. Clark<sup>4</sup>, Khai Luong<sup>4</sup>, Yi Song<sup>4</sup>, Yu-Chih Tsai<sup>4</sup>, Matthew Boitano<sup>4</sup>, Jyoti Gupta<sup>2</sup>, Shelise Y. Brooks<sup>2</sup>, Brian Schmidt<sup>2</sup>, Alice C. Young<sup>2</sup>, James W. Thomas<sup>2</sup>, Gerard G. Bouffard<sup>2</sup>, Robert W. Blakesley<sup>2</sup>, NISC Comparative Sequencing Program<sup>2</sup>, James C. Mullikin<sup>2</sup>, Jonas Korlach<sup>4</sup>, David K. Henderson<sup>3</sup>, Karen M. Frank<sup>3,\*</sup>, Tara N. Palmore<sup>3,\*</sup>, and Julia A. Segre<sup>1,\*</sup>

<sup>1</sup>National Human Genome Research Institute, Bethesda, MD



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## Tracking a plasmid during outbreak of KPC+ isolates at NIH



- Wide array of plasmids with carbapenem resistance genes found in several *Enterobacteriaceae* spp.
- Horizontal transfer of plasmids in the hospital environment
- Difficult to pin down person to person transmission



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## Roadblocks



- Cost and Time
- Bioinformatics consideration: making sense of the data
- Genotype: phenotype correlation (expression)
- Significance of a genome within a specimen: clinical correlation
- Quality Assurance: Curating and maintaining reliable database
- Regulatory considerations
- Patient outcomes

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## Molecular Assays: Making the Difference?

- Summary
  - New technology can provide accurate diagnosis in a shorter time
  - Consultative microbiology could enhance understanding of the “lab report”
  - Consultation with clinical pathologist help in choice of platform
  - With novel pathogens, antimicrobial resistance and technology laboratory stewardship becomes an integral part of patient care
  - Applications of NGS in clinical microbiology is promising but need careful cost benefit evaluation depending on the application

NorthShore  
University HealthSystem

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# Molecular Diagnostics and its Role in Infection Prevention

Dr. Sanchita Das, University of Chicago

A Webber Training Teleclass

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## Thank You!



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September 20, 2018	<p><a href="#"><u>THE SILENT TSUNAMI OF AZOLE-RESISTANCE IN THE OPPORTUNISTIC FUNGUS ASPERGILLUS FUMIGATUS</u></a> Speaker: <b>Prof. Paul E. Verweij</b>, Radboud University Center of Expertise in Mycology, The Netherlands</p>
September 27, 2018	<p><a href="#"><u>CHLORHEXIDINE USE AND BACTERIAL RESISTANCE</u></a> Speaker: <b>Prof. Jean Yves Maillard</b>, Cardiff University, Wales</p>
September 30, 2018	<p><i>(FREE European Teleclass - Broadcast live from the 2018 IPS conference)</i> <a href="#"><u>Cottrell Lecture ... SURVEILLANCE BY OBJECTIVES: USING MEASUREMENT IN THE PREVENTION OF HEALTHCARE ASSOCIATED INFECTIONS</u></a> Speaker: <b>Prof. Jennie Wilson</b>, University of West London</p>
October 2, 2018	<p><i>(FREE European Teleclass - Broadcast live from the 2018 IPS conference)</i> <a href="#"><u>Ayliffe Lecture ... (TO BE POSTED)</u></a> Speaker: <b>Prof. Shaheen Mehtar</b>, Stellenbosch University, Cape Town, South Africa</p>
October 11, 2018	<p><i>(FREE CBIC Teleclass)</i> <a href="#"><u>INFECTION CONTROL CHAMPIONS ARE MADE, NOT BORN</u></a></p>

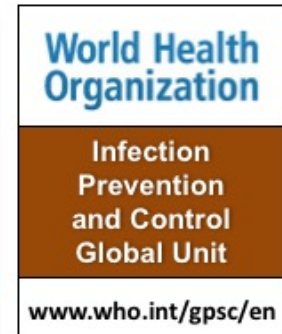
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